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CytoTune™-iPS 2.0 Sendai Reprogramming Kit

USER GUIDE

For efficient, integration-free reprogramming of somatic cells into induced pluripotent stem cells (iPSC)

Catalog Numbers A16517, A16518

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Revision 2.0

ThermoFisher SCIENTIFIC

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1.0	08 October 2013	New document

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Table of Contents

Product description

The CytoTune™-iPS 2.0 Sendai Reprogramming Kit contains three CytoTune™ 2.0 reprogramming vectors that are used for delivering and expressing key genetic factors necessary for reprogramming somatic cells into iPSCs.

Important! CytoTune™ 2.0 reprogramming vectors are **not** compatible with the reprogramming vectors from the original CytoTune™-iPS Reprogramming Kits (Cat. No. A13780-01, A13780-02). Do **not** mix or substitute CytoTune™ 2.0 reprogramming vectors with the reprogramming vectors from the original kits.

Important! This product must be used under Biosafety Level 2 (BL-2) containment with biological safety cabinet and laminar flow hood, and with appropriate personal safety equipment to prevent mucosal exposure/splash. For more information on BL-2 guidelines, see page 8.

Contents and storage

Component[1]	Cap color	Amount[2]		Storage
		A16517	A16518	
CytoTune™ 2.0 KOS	clear	100 µL	3 × 100 µL	–80°C
CytoTune™ 2.0 hc-Myc	white	100 µL	3 × 100 µL	
CytoTune™ 2.0 hKlf4	red	100 µL	3 × 100 µL	

[1]The titer of each CytoTune™ 2.0 reprogramming vector is lot-dependent. For the specific titer of your vectors, refer to the Certificate of Analysis (CoA) available on our website. Go to **www.thermofisher.com/cytotune** and search for the CoA by product lot number, which is printed on the vial.

[2]Each vial containing 100 µL of one of the CytoTune™ 2.0 reprogramming vector at a concentration of ≥ 8×10^7 cell infectious units/mL (CIU/mL).

Description of the system

Induced pluripotent stem cells (iPSC)

Induced pluripotent stem cells (iPSCs) are genetically reprogrammed adult cells which exhibit a pluripotent stem cell-like state similar to embryonic stem cells While these artificially generated cells are not known to exist in the human body, they show qualities remarkably similar to those of embryonic stem cells (ESC); thus, they are an invaluable new source of pluripotent cells for drug discovery, cell therapy, and basic research.

There are multiple methods to generate iPSCs, including retrovirus-mediated gene transduction and chemical induction. While retroviral vectors require integration into host chromosomes to express reprogramming genes, DNA-based vectors such as adenovirus, adeno-associated virus, and plasmid vectors exist episomally and do not require integration; however, they may still be integrated into host chromosomes at certain frequencies. Unlike these vectors, the CytoTune™ 2.0 reprogramming vectors do not integrate into the host genome or alter the genetic information of the host cell.

CytoTune™-iPS 2.0 Reprogramming System

CytoTune™-iPS 2.0 Reprogramming System uses vectors based on a modified, non-transmissible form of Sendai virus (SeV) to safely and effectively deliver and express key genetic factors necessary for reprogramming somatic cells into iPSCs. In contrast to many available protocols, which rely on viral vectors that integrate into the genome of the host cell, the CytoTune™-iPS 2.0 Reprogramming System uses vectors that are non-integrating and remain in the cytoplasm (i.e., they are zero-footprint). In addition, the host cell can be cleared of the vectors and reprogramming factor genes by exploiting the cytoplasmic nature of SeV and the functional temperature sensitivity mutations introduced into the key viral proteins.

The CytoTune™-iPS 2.0 Sendai Reprogramming Kit contains three SeV-based reprogramming vectors, and are optimized for generating iPSCs from human somatic cells. The reprogramming vectors in this kit have been engineered to increase biological and environmental safety (see **Safety features of the system**, page 8).

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Sendai virus

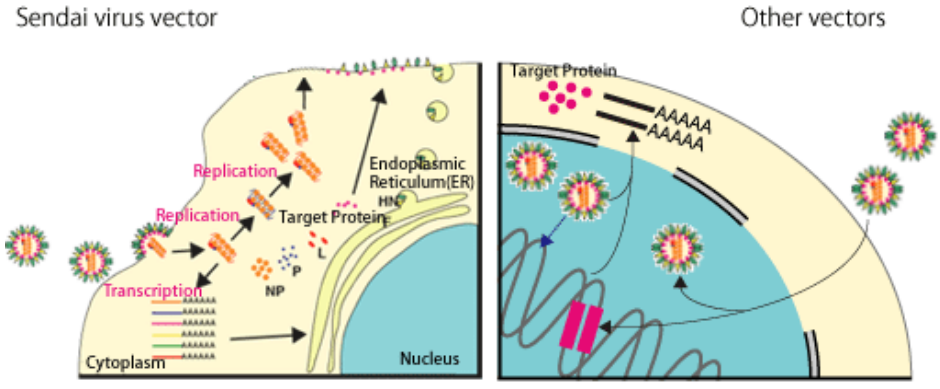
Sendai virus (SeV) is a respiratory virus of mouse and rat, classified as mouse parainfluenza virus type I belonging to the *Paramyxoviridae* family. SeV was first isolated in Japan in the early 1950s and is also called Hemagglutinating Virus of Japan (HVJ). SeV is an enveloped virus of 150–250 nm in diameter whose genome is a single chain RNA (15,384 bases) in the minus sense. Six genes coding for viral proteins are situated sequentially on the genome of the wild-type SeV in the following order (starting from the 3′ end):

- Nucleocapsid protein (NP) forms the core nucleocapsid complex with the genome RNA.
- Phosphoprotein (P) is the small subunit of the RNA polymerase.
- Matrix protein (M) supports the envelope structure from the inside.
- Fusion protein (F) fuses the viral envelope with cell membrane when the virus enters the cell.

  Note: The gene encoding the F protein is deleted from the CytoTune™ 2.0 reprogramming vectors, rendering them incapable of producing infectious particles from infected cells (see page 8).
- Hemagglutinin-Neuraminidase (HN) recognizes the cell surface receptor, sialic acid.
- Large protein (L) is the large subunit of RNA polymerase.

Because SeV infects cells by attaching itself to the sialic acid receptor present on the surface of many different cells, it can infect a wide range of cell types of various animal species. Activation of F protein by a protease is required for the virus-cell fusion process to take place. After infection, the virus goes through genome replication and protein synthesis, and then daughter virus particles are assembled and released.

Figure 1 Comparison of the lifecycles of non-integrating SeV vectors and other, integrating vectors



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CytoTune™ 2.0 reprogramming vectors

The table below lists the CytoTune™ 2.0 reprogramming vectors included in the CytoTune™-iPS 2.0 Sendai Reprogramming Kit. The reprogramming vectors include the four Yamanaka factors, Oct, Sox2, Klf4, and c-Myc, shown to be sufficient for efficient reprogramming.

CytoTune™ Sendai vector	Cap color	Factor	GenBank ID
CytoTune™ 2.0 KOS	clear	Human Klf4 Human Oct3/4 Human Sox2	BC029923.1 NM_002701.4 NM_003106.2
CytoTune™ 2.0 hc-Myc	white	Human c-Myc	K02276.1
CytoTune™ 2.0 hKlf4	red	Human Klf4	BC029923.1

Advantages of CytoTune™-iPS 2.0 Sendai Reprogramming Kit

- No genotoxicity: CytoTune™ 2.0 Sendai reprogramming vectors do not integrate into chromosomes of the target cells and potentially disrupt important genes.
- Wide range of targets: CytoTune™ 2.0 Sendai reprogramming vectors are capable of transducing a wide range of cell types in proliferative and quiescent states.
- High transduction efficiency with low multiplicity of infection (MOI).
- Short contact time of virus with target cells is sufficient to establish transduction.
- High level of expression of the transgenes.
- Fast expression of the transgenes: expression is detectable as early as 6–10 hours after transduction, with maximum expression detected more than 24 hours after transduction.
- Zero footprint: the vectors and transgenes can be eliminated from the cells.
- No production of infectious particles by the transduced cells.
- Derived from a virus that is non-pathogenic to humans.

Safety features of the system

Sendai virus (SeV) safety information

Host species: The host species for the Sendai virus (SeV) reported so far are mouse, rat, hamster, and guinea pigs, all of which have been described to be serologically positive.

Transmission: SeV is transmitted by aerosol and contact with respiratory secretions. The virus is highly contagious, but the infection does not persist in immunocompetent animals.

CytoTune™ 2.0 Sendai reprogramming vectors: CytoTune™ 2.0 Sendai reprogramming vectors in this kit are based on a modified, non-transmissible form of SeV, which has the Fusion protein (F) deleted, rendering the virus incapable of producing infectious particles from infected cells.

Inoculating animals with transduced cells: Although the CytoTune™ 2.0 Sendai reprogramming vectors are non-transmissible, cells that have been exposed to the virus should be tested with PCR or antibody staining to ensure the absence of the virus before being inoculated into animals. Animals that have already been infected with wild type SeV may be able to make infectious CytoTune™-2.0 Sendai virus.

Non-transmissible CytoTune™ 2.0 Sendai reprogramming vectors

SeV vectors used in this kit consist of viral proteins NP, P, M, F (activated), HN, and L, and the SeV genome RNA, from which the F gene is deleted. Because SeV infects cells by attaching itself to cell surface receptor sialic acid, present on the surface of many cell types of different species, the vectors are able to transduce a wide range of cells. However, they are no longer capable of producing infectious particles from infected cells, because the viral genome lacks the F-gene. In addition, the presence of functional mutations such as temperature sensitivity in the amino acid sequence of several SeV proteins (SeV/TSΔF, SeV/TS12ΔF, and SeV/TS15ΔF) renders the vectors easily removable from transduced cells.

Note: SeV vectors used in this kit were developed by DNAVEC Corporation (http://www.dnavec.co.jp) and their rights for commercial use are the property of DNAVEC Corporation.

Biosafety Level 2

Although human is not the natural host for the SeV, and the virus is non-pathogenic to humans, appropriate care must be taken to prevent the potential mucosal exposure to the virus. This product must be used under Biosafety Level 2 (BL-2) containment with biological safety cabinet and laminar flow hood, and with appropriate personal safety equipment to prevent mucosal exposure/splash. In the event that the virus comes into contact with skin or eyes, decontaminate by flushing with plenty of water and consult a physician. For more information on BL-2 guidelines, refer to *Biosafety in Microbiological and Biomedical Laboratories*, 5th ed., published by the Centers for Disease Control, which is available for downloading at: **www.cdc.gov/od/ohs/biosfty/bmbl5/bmbl5toc.htm**.

Guidelines for generating iPSCs

Experimental guidelines

- To maintain sterile culture conditions, carry out all of the procedures in this manual using sterile laboratory practices in a laminar flow hood.
- You can use the CytoTune™-iPS 2.0 Sendai Reprogramming Kit to reprogram a wide range of cell types in proliferative and quiescent states. However, the reprogramming efficiency may vary between different cell types (~0.01%–1%).
- For successful reprogramming, transduce your cells using all three reprogramming vectors.
  Note: For successful reprogramming, all four Yamanaka factors (i.e., Oct4, Sox2, Klf4, and c-Myc) need to be expressed in your host cell.
- Cells that have already been infected with Sendai virus are refractive to further infection by Sendai virus. Therefore, you cannot transduce cells with CytoTune™ 2.0 reprogramming vectors that have already been transduced with other Sendai vectors such as the CytoTune™-EmGFP Sendai Fluorescence Reporter or vice versa.
- One CytoTune™-iPS 2.0 Reprogramming Kit of three tubes supplies sufficient reagents to transduce a minimum of 1.5×10^6 cells at MOI=5-5-3 (i.e., KOS MOI=5, hc-Myc MOI=5, hKlf4 MOI=3).
- The titer of each CytoTune™ 2.0 Sendai reprogramming vector is lot-dependent. For the specific titer of your vectors, refer to the Certificate of Analysis (CoA) available on our website. Go to **thermofisher.com/cytotune** and search for the CoA by product lot number, which is printed on the vial.
- Viral titers can decrease dramatically with each freeze/thaw cycle. Avoid repeated freezing and thawing of your reprogramming vectors. Viral titer is not guaranteed for kits that have been refrozen or thawed.
- Prior to starting, ensure that the media are equilibrated to 37°C and appropriately gassed.

Positive control

For positive control, we recommend performing a reprogramming experiment with human neonatal foreskin fibroblast cells (strain BJ; ATCC no. CRL2522). Note that experimental conditions may vary among target cells and need to be optimized for each cell type. The example given in the following protocol does not guarantee the generation of iPSCs for all cell types.

Continued on next page

CytoTune™-EmGFP Sendai Fluorescence Reporter

The CytoTune™-EmGFP Sendai Fluorescence Reporter (Cat. No. A16519), available separately, is a fluorescent control vector carrying the Emerald Green Fluorescent Protein (EmGFP) gene. The fluorescent control vector allows you to determine whether your cell line of interest is amenable or refractive to transduction by the Sendai reprogramming vectors, including the vectors from the original CytoTune™-iPS Sendai Reprogramming Kits. We recommend testing your cell lines of interest using the CytoTune™-EmGFP Sendai Fluorescence Reporter before starting your reprogramming experiments.

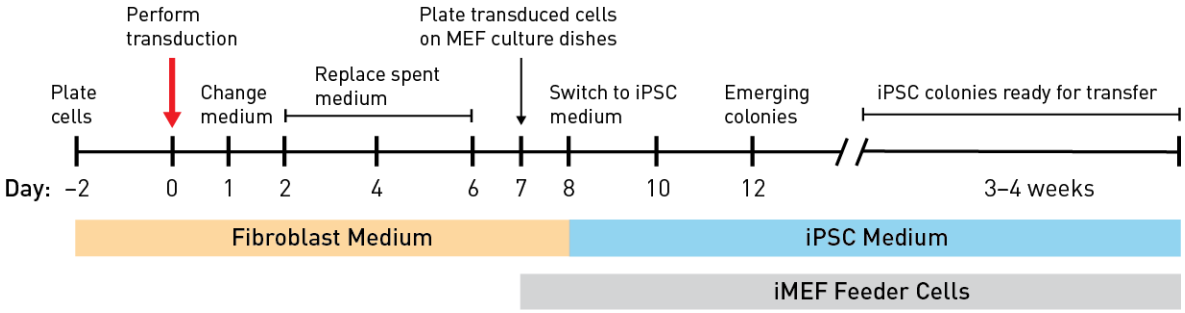
Note that you cannot transduce cells with CytoTune™ reprogramming vectors that have already been transduced with the CytoTune™-EmGFP Sendai Fluorescence Reporter or vice versa. If you wish to use the CytoTune™-EmGFP Sendai Fluorescence Reporter during reprogramming, you must add it to the cells at the same time as the reprogramming vectors.

For detailed instructions on using the CytoTune™-EmGFP Sendai Fluorescence Reporter, see page 60.

Experiment outline (Feeder-dependent)

Workflow

The major steps required for reprogramming human neonatal foreskin fibroblast cells using the CytoTune™-iPS 2.0 Sendai Reprogramming Kit to generate iPSCs cultured on MEF feeder-cells are shown below. Note that the timeline is provided as a guideline for experimental planning; actual timeline can vary based on the cell type and experimental conditions.



Reprogramming timeline

Day –2: Plate human fibroblasts with a passage number of 5 or lower into at least two wells of a 6-well plate in fibroblast medium so that they are 30–60% confluent on the day of transduction (Day 0).

Note: The kit supplies sufficient virus to transduce cells in at least 5 wells of a 6-well plate.

Day 0: Transduce the cells using the CytoTune™ 2.0 Sendai reprogramming vectors at the appropriate MOI. Incubate the cells overnight.

Day 1: Replace the medium with fresh complete fibroblast medium to remove the CytoTune™ 2.0 Sendai reprogramming vectors.

Day 2–6: Replace the spent medium every other day.

Day 5 or 6: Prepare MEF culture dishes.

Day 7: Plate transduced cells on MEF culture dishes in fibroblast medium.

Day 8: Change the medium to iPSC medium.

Day 9–28: Replace spent medium every day and monitor the culture vessels for the emergence of iPSC colonies. When iPSC colonies are ready for transfer, perform live staining, and pick and transfer undifferentiated iPSCs onto fresh MEF culture dishes for expansion.

Media for reprogramming fibroblasts (feeder-dependent)

For optimal reprogramming of human neonatal foreskin fibroblast cells using the CytoTune™-iPS 2.0 Sendai Reprogramming Kit to generate iPSCs cultured on MEF feeder-cells, use the following media at the designated stages of the reprogramming experiment:

- **Fibroblast medium** (page 52): Plating cells prior to transduction, expansion, post-transduction recovery of cells, plating of transduced cells on MEF culture dishes
- **iPSC medium** (page 53): Expansion of transduced cells on MEF culture dishes, live staining and picking of iPSCs

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Reprogram fibroblasts (Feeder-Dependent)

Required materials

Cells and vectors

- CytoTune™ 2.0 Sendai reprogramming vectors
  Note: For successful reprogramming, you need all three tubes of reprogramming vectors.
- Human fibroblast cells to reprogram
- *Optional*: Human neonatal foreskin fibroblast cells (strain BJ; ATCC no. CRL2522) as a positive reprogramming control
- Gibco™ Mouse Embryonic Fibroblasts (Irradiated) (Cat. No. S1520-100)

Media and reagents

- DMEM with GlutaMAX™-I (high glucose) (Cat. No. 10569-010)
- DMEM/F-12, GlutaMAX™ supplement (Cat. No. 12660-012)
- Fetal Bovine Serum (FBS), ES Cell-Qualified (Cat. No. 16141-079)
- KnockOut™ Serum Replacement (KSR) (Cat. No. 10565018)
- MEM Non-essential Amino Acids (NEAA) (Cat. No. 11140-050)
- Basic FGF, recombinant human (Cat. No. PHG0264)
- β-mercaptoethanol (Cat. No. 21985-023)
- *Optional:* Penicillin-Streptomycin, liquid (Cat. No. 15140-122)
- Attachment Factor (Cat. No. S-006-100)
- TrypLE™ Select Cell Dissociation Reagent (Cat. No. 12563) or 0.05% Trypsin/EDTA (Cat. No. 25300)
- Dulbecco's PBS (DPBS) without Calcium and Magnesium (Cat. No. 14190)

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Reprogram fibroblasts

The following protocol has been optimized to transduce one well of human neonatal foreskin fibroblast cells (strain BJ; ATCC no. CRL2522), as a positive control. We recommend that you optimize the protocol for your cell type, and add an appropriate number of conditions/wells to utilize the entire volume of virus.

Day –2: Prepare the cells for transduction

1. Two days before transduction, plate human neonatal foreskin fibroblast cells onto at least two wells of a 6-well plate at the appropriate density to achieve between 2×10^5–3×10^5 cells per well on the day of transduction (Day 0). One of the wells will be used to count cells for viral volume calculations.

   Note: Each CytoTune™-iPS 2.0 Sendai Reprogramming Kit supplies sufficient virus to transduce cells in at least 5 wells of a 6-well plate. We recommend using the entire volume of virus.

   Note: We recommend about 30–60% confluency on the day of transduction. Because overconfluency results in decreased transduction efficiency, we recommend replating your cells to achieve 30–60% confluency if your cells have become overconfluent during culturing.

2. Culture the cells for two more days, ensuring the cells have fully adhered and extended.

Day 0: Perform transduction

3. On the day of transduction, warm 1 mL of fibroblast medium in a water bath (see page 52 for recipe) for each well to be transduced.
4. Harvest the cells from **one well** to perform a cell count. **These cells will not be transduced**, but will be used to estimate the cell number in the other well(s) plated in Step 1.
5. Remove the cells from this well using 0.5 mL of TrypLE™ Select reagent or 0.05% trypsin/EDTA following the procedure recommended by the manufacturer and incubating at room temperature. When the cells have rounded up (1–3 minutes later), add 1 mL of fibroblast medium into each well, and collect the cells in a 15-mL conical centrifuge tube.
6. Count the cells using the desired method (e.g., Countess™ Automated Cell Counter), and calculate the volume of each virus needed to reach the target MOI using the **live cell count** and the titer information on the CoA.

$$\text{Volume of virus (µL)} = \frac{\text{MOI (CIU/cell)} \times \text{number of cells}}{\text{titer of virus (CIU/mL)} \times 10^{-3}\ \text{(mL/µL)}}$$

   Note: We recommend initially performing the transductions with MOIs of 5, 5, and 3 (i.e., KOS MOI=5, hc-Myc MOI=5, hKlf4 MOI=3). These MOIs can be optimized for your application.

   Note: The titer of each CytoTune™ 2.0 reprogramming vector is lot-dependent. For the specific titer of your vectors, go to **thermofisher.com/cytotune** and search for the CoA by product lot number, which is printed on the vial. Avoid re-freezing and thawing of the reprogramming vectors since viral titers can decrease dramatically with each freeze/thaw cycle.

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Reprogram fibroblasts, continued

7. Remove one set of CytoTune™ 2.0 Sendai tubes from the –80°C storage. Thaw each tube one at a time by first immersing the bottom of the tube in a 37°C water bath for 5–10 seconds, and then removing the tube from the water bath and allowing it to thaw at room temperature. Once thawed, briefly centrifuge the tube and place it immediately on ice.
8. Add the calculated volumes of each of the three CytoTune™ 2.0 Sendai tubes to 1 mL of fibroblast medium, pre-warmed to 37°C. Ensure that the solution is thoroughly mixed by pipetting the mixture gently up and down. Complete the next step within 5 minutes.
9. Aspirate the fibroblast medium from the cells, and add the reprogramming virus mixture prepared in Step 8 to the well containing the cells. Incubate the cells overnight in a 37°C incubator with a humidified atmosphere of 5% CO_2.

Day 1: Replace medium and culture cells

10. 24 hours after transduction, replace the medium with fresh fibroblast medium.

    Note: Depending on your cell type, you should expect to see some cytotoxicity 24–48 hours post-transduction, which can affect >50% of your cells. This is an indication of high uptake of the virus. We recommend that you continue culturing your cells and proceed with the protocol.

11. Culture the cells for 6 more days, changing the spent medium with fresh fibroblast medium every other day.

    Note: Depending on your cell type, you may observe high cell density before Day 5. We do **not** recommend passaging your cells onto MEF culture dishes before 7 days post-transduction. You may replace spent medium daily with fresh fibroblast medium if cultures become very dense.

Day 5 or 6: Prepare MEF culture dishes

12. One to two days before passaging the transduced fibroblasts onto MEF feeder-cells, prepare 6-well MEF culture dishes (see page 55).

Day 7: Plate transduced cells on MEF culture dishes

13. Seven days after transduction (Step 6, page 13), fibroblast cells are ready to be harvested and plated on MEF culture dishes. Remove the medium from the fibroblasts, and wash cells once with D-PBS.
14. To remove the cells from the 6-well plate, use 0.5 mL of TrypLE™ Select reagent or 0.05% trypsin/EDTA following the procedure recommended by the manufacturer and incubate at room temperature. When the cells have rounded up (1–3 minutes later), add 2 mL of fibroblast medium into each well, and collect the cells in a 15-mL conical centrifuge tube.

    Note: Because the cells can be very sensitive to trypsin at this point, minimize trypsin exposure time and incubate the cells at room temperature.

15. Centrifuge the cells at 200 × g for 4 minutes, aspirate the medium, and re-suspend the cells in an appropriate amount of fibroblast medium.

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Reprogram fibroblasts, continued

16. Count the cells using the desired method (e.g., Countess™ Automated Cell Counter), and seed the MEF culture dishes with 2×10^4–1×10^5 cells per well and incubate overnight in a 37°C incubator with a humidified atmosphere of 5% CO_2.

    Note: Reprogramming efficiency can vary widely, so we recommend plating at least two to four different densities. Depending on your cell type, you may need to plate most of your cells in the same well to ensure sufficient numbers of colonies.

    Note: It is strongly recommended to set aside cells at this point for RNA extraction to be used as a positive control in the RT-PCR or qPCR detection of the CytoTune™ vectors (see page 49). It is very important to include this positive control when performing detection of the CytoTune™ vectors.

Day 8 to 28: Feed and monitor the cells

17. 24 hours later, change the medium to iPSC medium (see page 53 for recipe), and replace the spent medium every day thereafter.
18. Starting on Day 8, observe the plates every other day under a microscope for the emergence of cell clumps indicative of reprogrammed cells (see **Visual identification**, page 45).

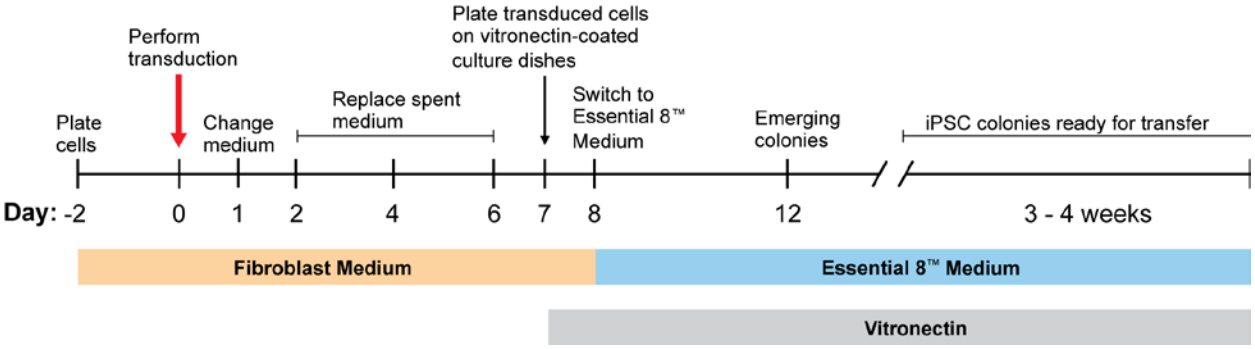
    Note: For BJ fibroblasts, we normally observe colony formation on Day 12 post-transduction. However, depending on your cell type, you may need to culture for up to 4 weeks before seeing colonies.
19. Three to four weeks after transduction, colonies should have grown to an appropriate size for transfer. The day before transferring the colonies, prepare MEF culture plates using Attachment Factor-coated 6- or 12-well plates.

    Note: We typically harvest colonies closer to three weeks to avoid differentiation.
20. When colonies are ready for transfer, perform live staining using Tra1-60 or Tra1-81 for selecting reprogrammed colonies (see **Live stain**, page 46).
21. Manually pick colonies and transfer them onto MEF plates (see **Pick iPSC colonies**, page 47).

Experiment outline (Feeder-Free)

Workflow

The major steps required for reprogramming human neonatal foreskin fibroblast cells using the CytoTune™-iPS 2.0 Sendai Reprogramming Kit to generate iPSCs cultured feeder-free on vitronectin-coated culture dishes are shown below. Note that the timeline is provided as a guideline for experimental planning; actual timeline can vary based on the cell type and experimental conditions.



Reprogramming timeline

Day –2: Plate human fibroblasts into at least two wells of a 6-well plate in fibroblast medium so that they are 30–60% confluent on the day of transduction (Day 0).

Note: The kit supplies sufficient virus to transduce cells in at least 5 wells of a 6-well plate.

Day 0: Transduce the cells using the CytoTune™ 2.0 Sendai reprogramming vectors at the appropriate MOI. Incubate the cells overnight.

Day 1: Replace the medium with fresh complete fibroblast medium to remove the CytoTune™ 2.0 Sendai reprogramming vectors.

Day 2–6: Replace the spent medium every other day.

Day 7: Plate transduced cells on vitronectin-coated culture dishes in fibroblast medium.

Day 8: Change the medium to complete Essential 8™ Medium.

Day 9–28: Replace spent medium every day and monitor the culture vessels for the emergence of iPSC colonies. When iPSC colonies are ready for transfer, perform live staining, and pick and transfer undifferentiated iPSCs onto fresh culture dishes for expansion.

Media for reprogramming fibroblasts (feeder-free)

For optimal reprogramming of human neonatal foreskin fibroblast cells using the CytoTune™-iPS 2.0 Sendai Reprogramming Kit to generate iPSCs cultured feeder free on vitronectin-coated culture dishes, use the following media at the designated stages of the reprogramming experiment:

- **Fibroblast medium** (page 52): Plating cells prior to transduction, expansion, post-transduction recovery of cells, plating of transduced cells on vitronectin-coated culture dishes
- **Complete Essential 8™ Medium** (page 53): Expansion of transduced cells on vitronectin-coated culture dishes, live staining and picking of iPSCs

Continued on next page

Reprogram fibroblasts (Feeder-Free)

Required materials

Cells and vectors

- CytoTune™ 2.0 Sendai reprogramming vectors
  Note: For successful reprogramming, you need all three tubes of reprogramming vectors.
- Human fibroblast cells to reprogram
- *Optional*: Human neonatal foreskin fibroblast cells (strain BJ; ATCC no. CRL2522) as a positive reprogramming control

Media and reagents

- DMEM with GlutaMAX™-I (high glucose) (Cat. No. 10569-010)
- Fetal Bovine Serum (FBS), ES Cell-Qualified (Cat. No. 16141-079)
- *Optional*: Penicillin-Streptomycin, liquid (Cat. No. 15140-122)
- TrypLE™ Select Cell Dissociation Reagent (Cat. No. 12563) or 0.05% Trypsin/EDTA (Cat. No. 25300)
- Dulbecco's PBS (DPBS) without Calcium and Magnesium (Cat. No. 14190)
- Essential 8™ Medium (Cat. No. A1517001)
- Vitronectin, truncated recombinant human (VTN-N) (Cat. No. A14700)

Continued on next page

Reprogram fibroblasts

The following protocol has been optimized to transduce one well of human neonatal foreskin fibroblast cells (strain BJ; ATCC no. CRL2522), as a positive control. We recommend that you optimize the protocol for your cell type, and add an appropriate number of conditions/wells to utilize the entire volume of virus.

Day –2: Prepare the cells for transduction

1. Two days before transduction, plate human neonatal foreskin fibroblast cells onto at least two wells of a 6-well plate at the appropriate density to achieve between 2×10^5–3×10^5 cells per well on the day of transduction (Day 0). One of the wells will be used to count cells for viral volume calculations.

   Note: Each CytoTune™-iPS 2.0 Sendai Reprogramming Kit supplies sufficient virus to transduce cells in at least 5 wells of a 6-well plate. We recommend using the entire volume of virus.

   Note: We recommend about 30–60% confluency on the day of transduction. Because overconfluency results in decreased transduction efficiency, we recommend replating your cells to achieve 30–60% confluency if your cells have become overconfluent during culturing.

2. Culture the cells for two more days, ensuring the cells have fully adhered and extended.

Day 0: Perform transduction

3. On the day of transduction, warm 1 mL of fibroblast medium in a water bath (see page 52 for recipe) for each well to be transduced.
4. Harvest the cells from **one well** to perform a cell count. **These cells will not be transduced**, but will be used to estimate the cell number in the other well(s) plated in Step 1.
5. Remove the cells from the 6-well plate using 0.5 mL of TrypLE™ Select reagent or 0.05% trypsin/EDTA following the procedure recommended by the manufacturer and incubating at room temperature. When the cells have rounded up (1–3 minutes later), add 1 mL of fibroblast medium into each well, and collect the cells in a 15-mL conical centrifuge tube.
6. Count the cells using the desired method (e.g., Countess™ Automated Cell Counter), and calculate the volume of each virus needed to reach the target MOI using the **live cell count** and the titer information on the CoA.

$$\text{Volume of virus (µL)} = \frac{\text{MOI (CIU/cell)} \times \text{number of cells}}{\text{titer of virus (CIU/mL)} \times 10^{-3}\ \text{(mL/µL)}}$$

   Note: We recommend initially performing the transductions with MOIs of 5, 5, and 3 (i.e., KOS MOI=5, hc-Myc MOI=5, hKlf4 MOI=3). These MOIs can be optimized for your application.

   Note: The titer of each CytoTune™ 2.0 reprogramming vector is lot-dependent. For the specific titer of your vectors, go to **thermofisher.com/cytotune** and search for the CoA by product lot number, which is printed on the vial. Avoid re-freezing and thawing of the reprogramming vectors since viral titers can decrease dramatically with each freeze/thaw cycle.

Continued on next page

Reprogram fibroblasts, continued

7. Remove one set of CytoTune™ 2.0 Sendai tubes from the –80°C storage. Thaw each tube one at a time by first immersing the bottom of the tube in a 37°C water bath for 5–10 seconds, and then removing the tube from the water bath and allowing it to thaw at room temperature. Once thawed, briefly centrifuge the tube and place it immediately on ice.
8. Add the calculated volumes of each of the three CytoTune™ 2.0 Sendai tubes to 1 mL of fibroblast medium, pre-warmed to 37°C. Ensure that the solution is thoroughly mixed by pipetting the mixture gently up and down. Complete the next step within 5 minutes.
9. Aspirate the fibroblast medium from the cells, and add the reprogramming virus mixture prepared in Step 8 to the well containing the cells. Incubate the cells overnight in a 37°C incubator with a humidified atmosphere of 5% CO_2.

Day 1: Replace medium and culture cells

10. 24 hours after transduction, replace the medium with fresh fibroblast medium.

    Note: Depending on your cell type, you should expect to see some cytotoxicity 24–48 hours post-transduction, which can affect >50% of your cells. This is an indication of high uptake of the virus. We recommend that you continue culturing your cells and proceed with the protocol.
11. Culture the cells for 6 more days, changing the spent medium with fresh fibroblast medium every other day.

    Note: Depending on your cell type, you may observe high cell density before Day 5. We do **not** recommend passaging your cells before 7 days post-transduction. You may replace spent medium daily with fresh fibroblast medium if cultures become very dense.

Day 7: Plate transduced cells on vitronectin-coated culture dishes

12. Coat a sufficient number of tissue culture dishes (e.g. 6-well, 60-mm, or 100-mm) with vitronectin (see page 57 for coating protocol).

    Note: Geltrex™ Membrane Matrix can be substituted for vitronectin; see page 57 for coating protocol.
13. Seven days after transduction (Step 9, above), fibroblast cells are ready to be harvested and plated on vitronectin-coated culture dishes. Remove the medium from the fibroblasts, and wash cells once with D-PBS.
14. To remove the cells from the 6-well plate, use 0.5 mL of TrypLE™ Select reagent or 0.05% trypsin/EDTA following the procedure recommended by the manufacturer and incubate at room temperature. When the cells have rounded up (1–3 minutes later), add 2 mL of fibroblast medium into each well, and collect the cells in a 15-mL conical centrifuge tube.

    Note: Because the cells can be very sensitive to trypsin at this point, minimize trypsin exposure time and incubate the cells at room temperature.
15. Centrifuge the cells at 200 × g for 4 minutes, aspirate the medium, and re-suspend the cells in an appropriate amount of fibroblast medium.

Continued on next page

Reprogram fibroblasts, continued

16. Count the cells using the desired method (e.g., Countess™ Automated Cell Counter), and seed the vitronectin-coated culture dishes with 2×10^4–1×10^5 cells per well of a 6-well and incubate overnight in a 37°C incubator with a humidified atmosphere of 5% CO_2.

    Note: Reprogramming efficiencies will typically be lower when using feeder-free conditions, so the number of cells plated should be increased accordingly. We recommend plating at least two to four different densities per well. Depending on your cell type, you may need to plate most of your cells on the same plate to ensure sufficient numbers of colonies.

    Note: It is strongly recommended to set aside cells at this point for RNA extraction to be used as a positive control in the RT-PCR or qPCR detection of the CytoTune™ vectors (see page 49). It is very important to include this positive control when performing detection of the CytoTune™ vectors.

Day 8 to 28: Feed and monitor the cells

17. 24 hours later, change the medium to complete Essential 8™ Medium (see page 53), and replace the spent medium every day thereafter.
18. Starting on Day 8, observe the plates every other day under a microscope for the emergence of cell clumps indicative of reprogrammed cells.

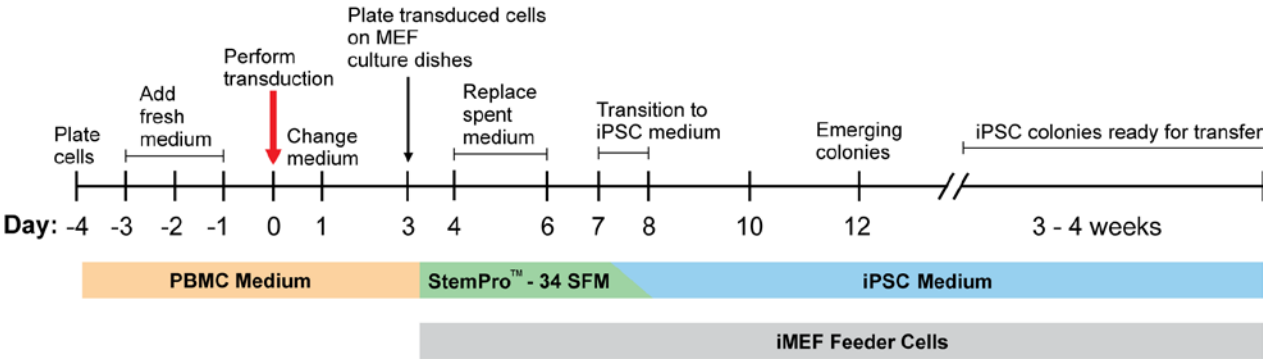
    Note: For BJ fibroblasts, we normally observe colony formation on Day 12 post-transduction. However, depending on your cell type, you may need to culture for up to 4 weeks before seeing colonies.
19. Three to four weeks after transduction, colonies should have grown to an appropriate size for transfer. When the colonies are ready for transfer, perform live staining using Tra1-60 or Tra1-81 for selecting reprogrammed colonies (see **Live stain**, page 46).

    Note: We typically harvest colonies closer to three weeks to avoid differentiation.
20. Manually pick undifferentiated iPSC colonies (see **Pick iPSC colonies**, page 47) and transfer them onto prepared vitronectin-coated 6- or 12-well culture plates for further expansion or analysis.

Experiment outline (Feeder-Dependent)

Workflow

The major steps required for reprogramming peripheral blood mononuclear cells (PBMCs) using the CytoTune™-iPS 2.0 Sendai Reprogramming Kit to generate iPSCs cultured on MEF feeder-cells are shown below. Note that the timeline is provided as a guideline for experimental planning; actual timeline can vary based on the cell type and experimental conditions.



Reprogramming timeline

Day –4: Plate peripheral blood mononuclear cells (PBMCs) at 5×10^5 cells/mL to the middle section of a 24-well plate in complete PBMC medium.

Day –3 to –1: Replace half of the medium with 0.5 mL of fresh complete PBMC medium.

Day 0: Transduce the cells using the CytoTune™ 2.0 Sendai reprogramming vectors at the appropriate MOI. Incubate the cells overnight.

Day 1: Replace the medium with fresh complete PBMC medium to remove the CytoTune™ 2.0 Sendai reprogramming vectors. Prepare MEF culture dishes for use on Day 3.

Day 3: Plate the transduced cells on MEF culture dishes in complete StemPro™-34 medium without cytokines.

Day 4–6: Replace spent complete StemPro™-34 medium without cytokines every other day.

Day 7: Start transitioning into iPSC medium by replacing half of the StemPro™-34 medium without cytokines with complete iPSC medium.

Day 8: Replace the entire medium with complete iPSC medium to conclude the transitioning, and continue culturing cells on MEF culture dishes

Day 9–28: Replace spent medium with fresh complete iPSC medium every day and monitor the culture vessels for the emergence of iPSC colonies. When iPSC colonies are ready for transfer, perform live staining, and pick and transfer undifferentiated iPSCs onto fresh MEF culture dishes for expansion.

Continued on next page

Reprogram peripheral blood mononuclear cells (PBMCs) (Feeder-Dependent)

Media for reprogramming PBMCs (feeder-dependent)

For optimal reprogramming of PBMCs using the CytoTune™-iPS 2.0 Sendai Reprogramming Kit to generate iPSCs cultured on MEF feeder-cells, use the following media at the designated stages of the reprogramming experiment:

- **PBMC medium** (page 53): Plating cells prior to transduction, expansion, post-transduction recovery of cells
- **StemPro™-34 medium without cytokines** (page 54): Plating of transduced cells on MEF culture dishes
- **iPSC medium** (page 53): Expansion of transduced cells on MEF culture dishes, live staining and picking of iPSCs

Required materials

Cells and vectors

- CytoTune™ 2.0 Sendai reprogramming vectors
  Note: For successful reprogramming, you need all three tubes of reprogramming vectors.
- Peripheral blood mononuclear cells (PBMCs) to reprogram
  Note: You can use PBMCs extracted from fresh blood by a conventional method (i.e., Ficoll-Paque purification) or frozen PBMCs.
- *Optional*: Human neonatal foreskin fibroblast cells (strain BJ; ATCC no. CRL2522) as a positive reprogramming control
  Note: If you are using this as a control, follow the protocol for reprogramming fibroblasts within this manual (page 12).
- Gibco™ Mouse Embryonic Fibroblasts (Irradiated) (Cat. No. S1520-100)

Media and reagents

- StemPro™-34 SFM Medium (Cat. No. 10639-011)
- L-Glutamine (Cat. No. 25030)
- DMEM with GlutaMAX™-I (High Glucose) (Cat. No. 10569-010)
- DMEM/F-12, GlutaMAX™ supplement (Cat. No. 10565018)
- Fetal Bovine Serum (FBS), ES Cell-Qualified (Cat. No. 16141-079)
- KnockOut™ Serum Replacement (KSR) (Cat. No. 10828-028)
- MEM Non-Essential Amino Acids (NEAA) (Cat. No. 11140-050)
- Basic FGF, Recombinant Human (Cat. No. PHG0264)
- SCF (C-Kit Ligand), Recombinant Human (Cat. No. PHC2111)
- FLT-3 Ligand, Recombinant Human (Cat. No. PHC9414)
- IL-3, Recombinant Human (Cat. No. PHC 0034)
- IL-6, Recombinant Human (Cat. No. PHC0065)
- β-Mercaptoethanol, 55 mM (Cat. No. 21985-023)
- *Optional*: Penicillin-Streptomycin, Liquid (Cat. No. 15140-122)
- *Optional*: Polybrene Hexadimethrine Bromide (Sigma Aldrich, Cat. No. H9268)
- Attachment Factor (Cat. No. S-006-100)
- Dulbecco's PBS (DPBS) without Calcium and Magnesium (Cat. No. 14190)
- Nunc™ 12-mL Round-Bottom Tubes (Cat. No. 150268)

Reprogram PBMCs

The following protocol has been optimized for peripheral blood mononuclear cells (PBMCs) isolated through density gradient centrifugation via Ficoll-Paque and frozen in FBS and DMSO-containing medium. We recommend that you optimize the protocol for your cell type.

Day –4: Seed PBMCs

1. Four days before transduction, remove vial(s) of PBMCs from liquid nitrogen storage. Thaw the vial quickly in 37°C water bath. When only a small ice crystal remains in the vial, remove it from the water bath. Spray the outside of the vial with 70% ethanol before placing it in the cell culture hood.
2. Gently transfer the PBMCs into a 15-mL conical tube. Slowly (drop-wise) add 5–10 mL pre-warmed complete PBMC medium (see page 53 for recipe) to the cell suspension. Remove an aliquot of cells to count and determine cell viability.

   Note: PBMC medium consists of complete StemPro™-34 medium containing the appropriate cytokines; aliquot the cytokines and add fresh daily.
3. Centrifuge the cell suspension at 200 × *g* for 10 minutes, discard the supernatant, and resuspend the cells in complete PBMC medium to 5×10^5 cells/mL.
4. Add 1 mL per well to the middle section of a 24-well plate to prevent excessive evaporation of the medium during incubation.

   Note: use at least 4 wells to ensure a sufficient number of cells on Day 0
5. Incubate the cells in a 37°C incubator with a humidified atmosphere of 5% CO_2.

Day –3 to –1: Observe cells and add fresh medium

6. Feed the cells daily, gently remove 0.5 mL of the medium from each well, and replace it with 0.5 mL of fresh complete PBMC medium, trying not to disturb the cells. If cells are present in 0.5 mL removed from the wells, centrifuge the cell suspension at 200 × *g* for 10 minutes, discard the supernatant, and resuspend the cells in 0.5 mL fresh PBMC medium before adding them back to the plate.

   Note: Some cell death is generally observed the first day after the thaw. Some cells may adhere to the surface of the tissue culture plate. Proceed with the cells in suspension, and leave behind any attached cells. Cells will not proliferate, but should maintain stable cell number for the first few days (PBMCs contain a variety of cells, and the current media system is only targeting a small population).

Continued on next page

Reprogram PBMCs, continued

Day 0: Count cells and perform transduction

7. Count the cells using the desired method (e.g., Countess™ Automated Cell Counter), and calculate the volume of each virus needed to reach the target MOI using the **live cell count** and the titer information on the CoA.

$$\text{Volume of virus (µL)} = \frac{\text{MOI (CIU/cell)} \times \text{number of cells}}{\text{titer of virus (CIU/mL)} \times 10^{-3}\ \text{(mL/µL)}}$$

Note: We recommend initially performing the transductions with MOIs of 5, 5, and 3 (i.e., KOS MOI=5, hc-Myc MOI=5, hKlf4 MOI=3). These MOIs can be optimized for your application.

Note: The titer of each CytoTune™ 2.0 reprogramming vector is lot-dependent. For the specific titer of your vectors, go to **thermofisher.com/cytotune** and search for the CoA by product lot number, which is printed on the vial. Avoid re-freezing and thawing of the reprogramming vectors since viral titers can decrease dramatically with each freeze/thaw cycle.

8. For each transduction, pipette 2.5 x 10^5–5 x 10^5 cells into a round bottom tube.
9. Remove CytoTune™ 2.0 Sendai tubes from the –80°C storage. Thaw each tube one at a time by first immersing the bottom of the tube in a 37°C water bath for 5–10 seconds, and then removing the tube from the water bath and allowing it to thaw at room temperature. Once thawed, briefly centrifuge the tube and place it immediately on ice.
10. Add the calculated volumes of each of the three CytoTune™ 2.0 Sendai tubes to 1 mL of PBMC medium, pre-warmed to 37°C. Ensure that the solution is thoroughly mixed by pipetting the mixture gently up and down. Complete the next step within 5 minutes.
11. Add the reprogramming virus mixture prepared in Step 10 to the round-bottom tube containing PBMCs prepared in Step 8. Total volume should now be between 1–1.5 mL. Place cap tightly onto the tube, and wrap with Parafilm. Centrifuge the cells and virus at 1000 x g for 30 minutes at room temperature. Once the centrifugation is complete, add an additional 1 mL of PBMC medium to the tube, re-suspend the cells, and transfer them to 1 well of a 12-well plate (total volume should now be between 2–2.5 mL). Incubate the plate overnight in a 37°C incubator with a humidified atmosphere of 5% CO_2.

    Note: Although this centrifugation step is not required, it significantly increases the transduction and reprogramming efficiencies. If the centrifugation step is omitted, transductions can be performed in a 24-well plate using 0.3 mL of total volume of cells, virus, and medium. Adding 4 µg/mL of Polybrene to the medium at the time of transduction may increase transduction efficiencies, but should only be used if the centrifugation step is not performed.

Day 1: Replace medium and culture cells

12. The next day, remove the cells and medium from the culture plate and transfer to a 15-mL centrifuge tube. Rinse the well gently with 1 mL of medium to ensure most of the cells are harvested.

Continued on next page

Reprogram PBMCs, continued

13. Remove the CytoTune™ 2.0 Sendai viruses by centrifuging the cell suspension at 200 × *g* for 10 minutes, aspirating the supernatant, and resuspending the cells in 0.5 mL of complete PBMC medium per well of a 24-well plate.

    Note: The cells may have drastic cell death (>60%); continue with the protocol using the **live** cell count. For the first 48 hours, observe the cells under the microscope for changes in cell morphology as a validation of transduction. Expect large, aggregated cells.

    Note: To prevent attachment of any cells prior to plating onto MEF, it may be beneficial to use a low attachment 24-well plate.

14. Culture the cells at 37°C in a humidified atmosphere of 5% CO_2 for 2 days. No media change is required during this time.

    Note: While the cells are incubating (1–2 days before plating the transduced cells), prepare MEF culture plates. You will need to have MEF feeder cells in at least two wells of a 6-well plate for each well of transduced cells (see page 55).

Day 3: Plate cells on MEF culture dishes

15. Count the cells using the desired method (e.g., Countess™ Automated Cell Counter) and seed the 6-well MEF culture plates with 1×10^4 –1×10^5 live cells per well in 2 mL of complete StemPro™-34 medium without the cytokines.

    Note: It may be necessary to plate more than two different densities, as the reprogramming efficiencies of PBMCs can vary widely between donors. If sufficient cells are available, it is recommended to plate 4–6 different densities, ranging from 1×10^4–1×10^5 cells per well of a 6-well plate.

    Note: It is strongly recommended to set aside cells at this point for RNA extraction to be used as a positive control in the RT-PCR or qPCR detection of the CytoTune™ vectors. It is very important to include this positive control when performing detection of the CytoTune™ vectors.

16. Incubate the cells at 37°C in a humidified atmosphere of 5% CO_2.

Day 4–6: Replace spent medium

17. Every other day, gently remove 1 mL (half) of the spent medium from the cells and replace it with 1 mL of fresh complete StemPro™-34 medium without cytokines and without disturbing cells.

    Note: The transduced PBMCs may only be loosely attached for the first few days after plating. Be sure to perform media changes gently during this time. If cells are present in 1 mL removed from the wells, centrifuge the cell suspension at 200 × g for 10 minutes, discard the supernatant, and resuspend the cells in 1 mL fresh PBMC medium before adding them back to the plate.

Day 7: Start transitioning cells to iPSC medium

18. Prepare 100 mL of complete iPSC medium (See page 53).
19. Remove 1 mL (half) of StemPro™-34 medium from the cells and replace it with 1 mL of iPSC medium to start the adaptation of the cells to the new culture medium.

Day 8 to 28: Feed and monitor the cells

20. 24 hours later (day 8), change the full volume of the medium to iPSC medium, and replace the spent medium every day thereafter.

    Note: plated PBMCs should have fully attached by this point. Any cells that have not attached will likely never do so.

21. Starting on day 8, observe the plates every other day under a microscope for the emergence of cell clumps indicative of reprogrammed cells (see Figure 2, page 26).
22. By day 15 to 21 after transduction, colonies should have grown to an appropriate

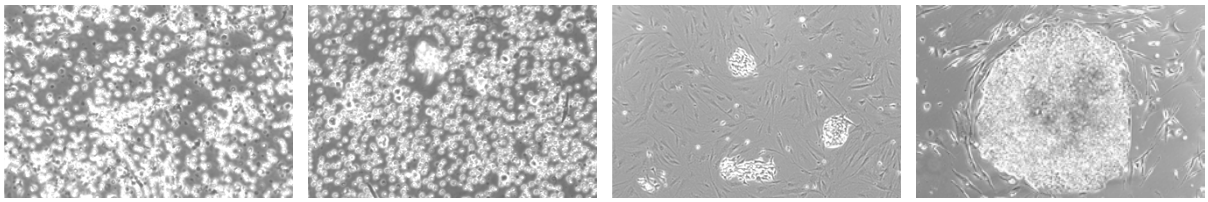
size for transfer. The day before transferring the colonies, prepare MEF culture plates using Attachment Factor-coated 6- or 12-well plates (see page 55).

Note: We typically harvest colonies closer to 3 weeks to avoid differentiation.

23. When colonies are ready for transfer, perform live staining using Tra1-60 or Tra1-81 for selecting reprogrammed colonies if desired (see **Live stain**, page 46).
24. Manually pick colonies and transfer them onto prepared MEF plates (see Pick iPSC colonies, page 47).

Expected results

Figure 2 Colony formation for iPSC generated from PBMC. Cells are cultured in complete PBMC medium (complete StemPro™-34 SFM + cytokines) for 4 days. On day 0 **(panel A)** cells are transduced overnight at an MOI of 5-5-3 (KOS MOI=5, hc-Myc MOI=5, hKlf4 MOI=3). At day 3 **(panel B)**, the cells show morphological changes indicating reprogramming and are plated on MEF feeder layers. The cells are allowed to proliferate on MEF feeder layers and colony formation is observed from day 8 **(panel C)** to day 14 **(panel D)**.

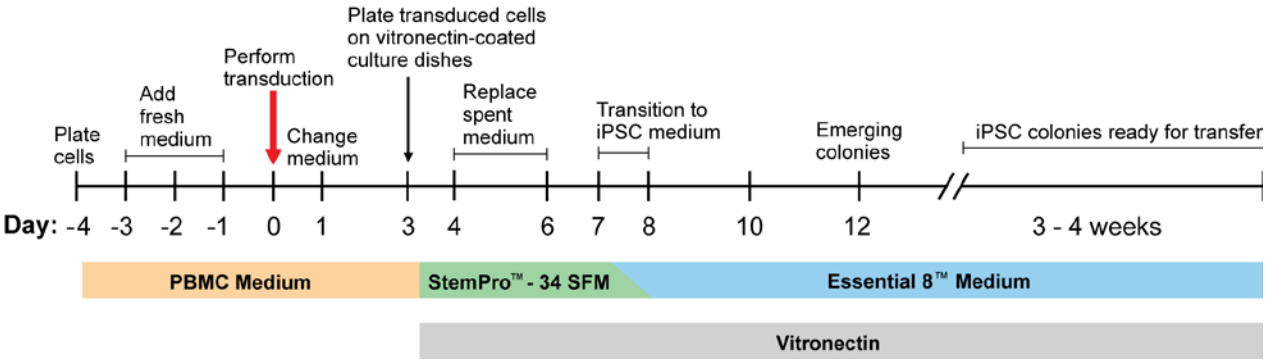


A **B** **C** **D**

Experiment outline (Feeder-Free)

Workflow

The major steps required for reprogramming peripheral blood mononuclear cells (PBMCs) using the CytoTune™-iPS 2.0 Sendai Reprogramming Kit to generate iPSCs cultured feeder-free on vitronectin-coated culture dishes are shown below. Note that the timeline is provided as a guideline for experimental planning; actual timeline can vary based on the cell type and experimental conditions.



Reprogramming timeline

Day –4: Plate peripheral blood mononuclear cells (PBMCs) at 5×10^5 cells/mL to the middle section of a 24-well plate in complete PBMC medium.

Day –3 to –1: Replace half of the medium with 0.5 mL of fresh complete PBMC medium.

Day 0: Transduce the cells using the CytoTune™ 2.0 Sendai reprogramming vectors at the appropriate MOI. Incubate the cells overnight.

Day 1: Replace the medium with fresh complete PBMC medium to remove the CytoTune™ 2.0 Sendai reprogramming vectors.

Day 3: Plate the transduced cells on vitronectin-coated culture dishes in complete StemPro™-34 medium without cytokines.

Day 4–6: Replace spent complete StemPro™-34 medium without cytokines every other day.

Day 7: Start transitioning into Essential 8™ Medium by replacing half of the StemPro™-34 medium without cytokines with Essential 8™ Medium.

Day 8: Replace the entire medium with Essential 8™ Medium to conclude the transitioning, and continue culturing cells on vitronectin-coated culture dishes

Day 9–28: Replace spent medium with fresh Essential 8™ Medium every day and monitor the culture vessels for the emergence of iPSC colonies. When iPSC colonies are ready for transfer, perform live staining, and pick and transfer undifferentiated iPSCs onto fresh vitronectin-coated culture dishes for expansion.

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Reprogram Peripheral Blood Mononuclear Cells (PBMCs) (Feeder-Free)

Media for reprogramming PBMCs (feeder-free)

For optimal reprogramming of PBMCs using the CytoTune™-iPS 2.0 Sendai Reprogramming Kit to generate iPSCs cultured on MEF feeder-cells, use the following media at the designated stages of the reprogramming experiment:

- **PBMC medium** (page 53): Plating cells prior to transduction, expansion, post-transduction recovery of cells
- **StemPro™-34 medium without cytokines** (page 54): Plating of transduced cells on MEF culture dishes
- **Complete Essential 8™ Medium** (page 53): Expansion of transduced cells on vitronectin-coated culture dishes, live staining and picking of iPSCs

Required materials

Cells and vectors

- CytoTune™ 2.0 Sendai reprogramming vectors
  Note: For successful reprogramming, you need all three tubes of reprogramming vectors.
- Peripheral blood mononuclear cells (PBMCs) to reprogram
  Note: You can use PBMCs extracted from fresh blood by a conventional method (i.e., Ficoll-Paque purification) or frozen PBMCs.
- *Optional*: Human neonatal foreskin fibroblast cells (strain BJ; ATCC no. CRL2522) as a positive reprogramming control
  Note: If you are using this as a control, follow the protocol for reprogramming fibroblasts within this manual (page 12).

Media and reagents

- StemPro™-34 SFM Medium (Cat. No. 10639-011)
- L-Glutamine (Cat. No. 25030)
- DMEM with GlutaMAX™-I (High Glucose) (Cat. No. 10569-010)
- KnockOut™ DMEM/F-12 (Cat. No. 12660-012)
- Fetal Bovine Serum (FBS), ES Cell-Qualified (Cat. No. 16141-079)
- SCF (C-Kit Ligand), Recombinant Human (Cat. No. PHC2111)
- FLT-3 Ligand, Recombinant Human (Cat. No. PHC9414)
- IL-3, Recombinant Human (Cat. No. PHC 0034)
- IL-6, Recombinant Human (Cat. No. PHC0065)
- *Optional*: Penicillin-Streptomycin, Liquid (Cat. No. 15140-122)
- *Optional*: Polybrene Hexadimethrine Bromide (Sigma Aldrich, Cat. No. H9268)
- Dulbecco's PBS (DPBS) without Calcium and Magnesium (Cat. No. 14190)
- Essential 8™ Medium (Cat. No. A1517001)
- Vitronectin, truncated recombinant human (VTN-N) (Cat. No. A14700)
- Nunc™ 14 mL Round-Bottom Tubes (Cat. No. 150268)

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Reprogramming protocol

The following protocol has been optimized for peripheral blood mononuclear cells (PBMCs) isolated through density gradient centrifugation via Ficoll-Paque and frozen in FBS and DMSO-containing medium. We recommend that you optimize the protocol for your cell type.

Day –4: Seed PBMCs

1. Four days before transduction, remove vial(s) of PBMCs from liquid nitrogen storage. Thaw the vial quickly in 37°C water bath. When only a small ice crystal remains in the vial, remove it from the water bath. Spray the outside of the vial with 70% ethanol before placing it in the cell culture hood.
2. Gently transfer the PBMCs into a 15-mL conical tube. Slowly (drop-wise) add 5–10 mL pre-warmed complete PBMC medium (see page 53 for recipe) to the cell suspension. Remove an aliquot of cells to count and determine cell viability.

   Note: PBMC medium consists of complete StemPro™-34 medium containing the appropriate cytokines; aliquot the cytokines and add fresh daily.
3. Centrifuge the cell suspension at 200 × *g* for 10 minutes, discard the supernatant, and resuspend the cells in complete PBMC medium to 5×10^5 cells/mL.
4. Add 1 mL per well to the middle section of a 24-well plate to prevent excessive evaporation of the medium during incubation.

   Note: Use at least 4 wells to ensure a sufficient number of cells on Day 0.
5. Incubate the cells in a 37°C incubator with a humidified atmosphere of 5% CO_2.

Day –3 to –1: Observe cells and add fresh medium

6. Feed the cells daily, gently remove 0.5 mL of the medium from each well, and replace it with 0.5 mL of fresh complete PBMC medium, trying not to disturb the cells. If cells are present in 0.5 mL removed from the wells, centrifuge the cell suspension at 200 × *g* for 10 minutes, discard the supernatant, and resuspend the cells in 0.5 mL fresh PBMC medium before adding them back to the plate.

   Note: Some cell death is generally observed the first day after the thaw. Some cells may adhere to the surface of the tissue culture plate. Proceed with the cells in suspension, and leave behind any attached cells. Cells will not proliferate, but should maintain stable cell number for the first few days (PBMCs contain a variety of cells, and the current media system is only targeting a small population).

Continued on next page

Reprogram PBMCs, continued

Day 0: Count cells and perform transduction

7. Count the cells using the desired method (e.g., Countess™ Automated Cell Counter), and calculate the volume of each virus needed to reach the target MOI using the **live cell count** and the titer information on the CoA.

$$\text{Volume of virus (µL)} = \frac{\text{MOI (CIU/cell)} \times \text{number of cells}}{\text{titer of virus (CIU/mL)} \times 10^{-3}\ \text{(mL/µL)}}$$

Note: We recommend initially performing the transductions with MOIs of 5, 5, and 3 (i.e., KOS MOI=5, hc-Myc MOI=5, hKlf4 MOI=3). These MOIs can be optimized for your application.

Note: The titer of each CytoTune™ 2.0 reprogramming vector is lot-dependent. For the specific titer of your vectors, go to **thermofisher.com/cytotune** and search for the CoA by product lot number, which is printed on the vial. Avoid re-freezing and thawing of the reprogramming vectors since viral titers can decrease dramatically with each freeze/thaw cycle.

8. For each transduction, pipette 2.5 x 10^5 –5 x 10^5 cells into a round bottom tube.
9. Remove CytoTune™ 2.0 Sendai tubes from the –80°C storage. Thaw each tube one at a time by first immersing the bottom of the tube in a 37°C water bath for 5–10 seconds, and then removing the tube from the water bath and allowing it to thaw at room temperature. Once thawed, briefly centrifuge the tube and place it immediately on ice.
10. Add the calculated volumes of each of the three CytoTune™ 2.0 Sendai tubes to 1 mL of PBMC medium, pre-warmed to 37°C. Ensure that the solution is thoroughly mixed by pipetting the mixture gently up and down. Complete the next step within 5 minutes.
11. Add the reprogramming virus mixture prepared in Step 10 to the round-bottom tube containing PBMCs prepared in Step 8. Total volume should now be between 1 –1.5 mL. Place cap tightly onto the tube, and wrap with Parafilm. Centrifuge the cells and virus at 1000 x g for 30 minutes at room temperature. Once the centrifugation is complete, add an additional 1 mL of PBMC medium to the tube, re-suspend the cells, and transfer them to 1 well of a 12-well plate (total volume should now be between 2 –2.5 mL). Incubate the plate overnight in a 37°C incubator with a humidified atmosphere of 5% CO_2.

    Note: Although this centrifugation step is not required, it significantly increases the transduction and reprogramming efficiencies. If the centrifugation step is omitted, transductions can be performed in a 24-well plate using 0.3 mL of total volume of cells, virus, and medium. Adding 4 µg/mL of Polybrene to the medium at the time of transduction may increase transduction efficiencies, but should only be used if the centrifugation step is not performed.

Day 1: Replace medium and culture cells

12. The next day, remove the cells and medium from the culture plate and transfer to a 15-mL centrifuge tube. Rinse the well gently with 1 mL of medium to ensure most of the cells are harvested.

Continued on next page

Reprogram PBMCs, continued

13. Remove the CytoTune™ 2.0 Sendai viruses by centrifuging the cell suspension at 200 × *g* for 10 minutes, aspirating the supernatant, and resuspending the cells in 0.5 mL of complete PBMC medium per well of a 24-well plate.

    Note: The cells may have drastic cell death (>60%); continue with the protocol using the **live** cell count. For the first 48 hours, observe the cells under the microscope for changes in cell morphology as a validation of transduction. Expect large, aggregated cells.

    Note: To prevent attachment of any cells prior to plating onto vitronectin, it may be beneficial to use a low attachment 24-well plate.

14. Culture the cells at 37°C in a humidified atmosphere of 5% CO_2 for 2 days. No media change is required during this time.

Day 3: Plate cells on vitronection-coated culture dishes

15. Coat a sufficient number of tissue culture dishes (e.g. 6-well, 60-mm, or 100-mm) with vitronectin (see page 57 for coating protocol).

    Note: Geltrex™ Membrane Matrix can be substituted for vitronectin; see page 57 for coating protocol.

16. Count the cells using the desired method (e.g., Countess™ Automated Cell Counter) and seed the 6-well vitronectin-coated culture plates with 1 x 10^4 - 1 x 10^5 live cells per well in 2 mL of complete StemPro™-34 medium without the cytokines.

    Note: It may be necessary to plate more than two different densities, as the reprogramming efficiencies of PBMCs can vary widely between donors. If sufficient cells are available, it is recommended to plate 4–6 different densities, ranging from 1 x 10^4–1 x 10^5 cells per well of a 6-well plate.

    Note: It is strongly recommended to set aside cells at this point for RNA extraction to be used as a positive control in the RT-PCR or qPCR detection of the CytoTune™ vectors. It is very important to include this positive control when performing detection of the CytoTune™ vectors.

17. Incubate the cells at 37°C in a humidified atmosphere of 5% CO_2.

Day 4–6: Replace spent medium

18. Every other day, gently remove 1 mL (half) of the spent medium from the cells and replace it with 1 mL of fresh complete StemPro™-34 medium without cytokines and without disturbing cells.

    Note: The transduced PBMCs may only be loosely attached for the first few days after plating. Be sure to perform media changes gently during this time. If cells are present in 1 mL removed from the wells, centrifuge the cell suspension at 200 × g for 10 minutes, discard the supernatant, and resuspend the cells in 1 mL fresh PBMC medium before adding them back to the plate.

Day 7: Start transitioning cells to Essential 8™ Medium

19. Prepare Essential 8™ Medium. See page 53.

20. Remove 1 mL (half) of StemPro™-34 medium from the cells and replace it with 1 mL of Essential 8™ Medium to start the adaptation of the cells to the new culture medium.

Day 8 to 28: Feed and monitor the cells

21. 24 hours later (day 8), change the full volume of the medium to Essential 8™ Medium, and replace the spent medium every day thereafter.

    Note: Plated PBMCs should have fully attached by this point. Any cells that have not attached will likely never do so.

22. Starting on day 8, observe the plates every other day under a microscope for the emergence of cell clumps indicative of reprogrammed cells (see Figure 2,

page 26).

23. By day 15 to 21 after transduction, colonies should have grown to an appropriate size for transfer.

    Note: We typically harvest colonies closer to 3 weeks to avoid differentiation.

24. When colonies are ready for transfer, perform live staining using Tra1-60 or Tra1-81 for selecting reprogrammed colonies if desired (see **Live stain**, page 46).

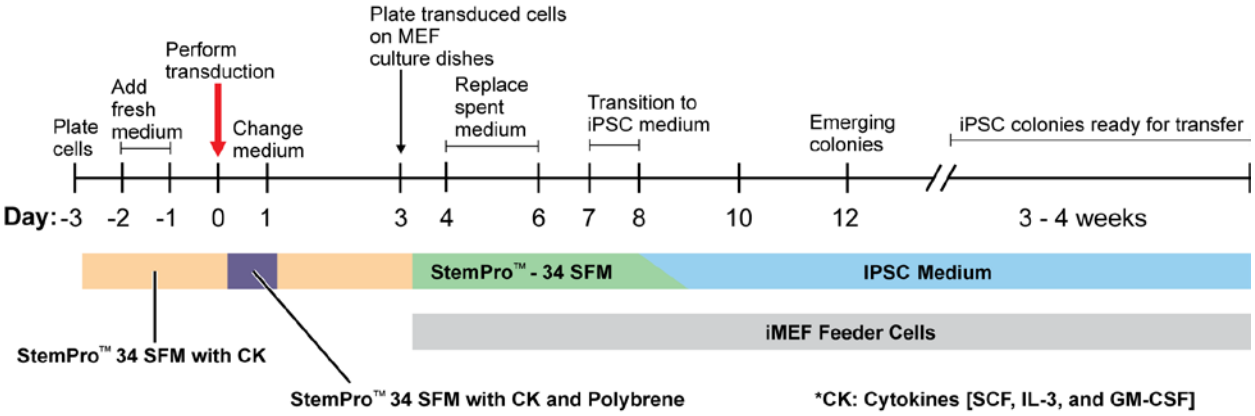
25. Manually pick colonies and transfer them onto prepared vitronectin-coated 6- or 12-well culture plates (see **Pick iPSC colonies**, page 47).

Experiment outline (Feeder-Dependent)

Workflow

The major steps required for reprogramming StemPro™ CD34+ cells using the CytoTune™-iPS 2.0 Sendai Reprogramming Kit to generate iPSCs cultured on MEF feeder-cells are shown below.

Note: The timeline is provided as a guideline for experimental planning; actual timeline can vary based on the cell type and experimental conditions.



Reprogramming timeline

Day –3: Plate StemPro™ CD34+ cells into at least two wells of a 24-well plate in complete StemPro™-34 medium containing cytokines (i.e., SCF, IL-3 and GM-CSF).

Day –1: Replace 0.5 mL of spent medium with 0.5 mL of fresh complete StemPro™-34 medium containing cytokines.

Day –2: Replace 0.5 mL of spent medium with 1 mL of fresh complete StemPro™-34 medium containing cytokines.

Day 0: Transduce the cells using the CytoTune™ 2.0 Sendai reprogramming vectors at the appropriate MOI in StemPro™-34 medium containing cytokines and 4 µg/mL of Polybrene. Incubate the cells overnight.

Day 1: Replace the medium with fresh StemPro™-34 medium containing cytokines (no Polybrene) to remove the CytoTune™ 2.0 Sendai reprogramming vectors. Prepare MEF culture dishes for use on Day 3.

Day 3: Plate the transduced cells on MEF culture dishes in complete StemPro™-34 medium without cytokines.

Day 4–6: Replace half of the spent medium with fresh complete StemPro™-34 medium without cytokines every other day.

Day 7: Start transitioning into iPSC medium by replacing half of the StemPro™-34 medium without cytokines with complete iPSC medium.

Day 8: Replace the entire medium with complete iPSC medium to conclude the transitioning and continue culturing cells on MEF culture dishes.

Day 9–28: Replace spent medium with fresh complete iPSC medium every day and monitor the culture vessels for the emergence of iPSC colonies. When iPSC colonies are ready for transfer, perform live staining, and pick and transfer undifferentiated iPSCs onto fresh MEF culture dishes for expansion.

Reprogram StemPro™ CD34+ Cells (Feeder-Dependent)

Media for reprogramming StemPro™ CD34+ cells (feeder-dependent)

For optimal reprogramming of CD34+ cells using the CytoTune™-iPS 2.0 Sendai Reprogramming Kit to generate iPSCs cultured on MEF feeder-cells, use the following media at the designated stages of the reprogramming experiment:

- **StemPro™-34 medium containing cytokines (i.e., SCF, IL-3 and GM-CSF)** (page 54): Plating cells prior to transduction, post-transduction recovery of cells
- **StemPro™-34 medium containing cytokines + 4 µg/mL of Polybrene**: Transduction
- **StemPro™-34 medium without cytokines** (page 54): Plating of transduced cells on MEF culture dishes
- **iPSC medium** (page 53): Expansion of transduced cells on MEF culture dishes, live staining and picking of iPSCs

Required materials

Cells and vectors

- CytoTune™ 2.0 Sendai reprogramming vectors
  Note: For successful reprogramming, you need all three tubes of reprogramming vectors.
- StemPro™ CD34+ cells to reprogram
  Note: StemPro™ CD34+ cells are available as part of the StemPro™-34 Medium and CD34+ Cell Kit (Cat. No. A14059).
- *Optional*: Human neonatal foreskin fibroblast cells (strain BJ; ATCC no. CRL2522) as a positive reprogramming control
  Note: If you are using this as a control, follow the protocol for reprogramming fibroblasts within this manual (page 12).
- Gibco™ Mouse Embryonic Fibroblasts (Irradiated) (Cat. No. S1520-100)

Media and reagents

- StemPro™-34 Medium and CD34+ Cell Kit (Cat. No. A14059)
- Recombinant Human SCF Lyophilized (Cat. No. PHC2111)
- Recombinant Human IL-3 Lyophilized (Cat. No. PHC0031)
- Recombinant Human GM-CSF Lyophilized (Cat. No. PHC2011)
- Dulbecco's Modified Eagle Medium (DMEM), High Glucose, with GlutaMAX™-I and Pyruvate (Cat. No. 10569-010)
- DMEM/F-12, GlutaMAX™ supplement (Cat. No. 10565018)
- Fetal Bovine Serum (FBS), ESC-Qualified, US Origin (Cat. No. 16141-079)
- KnockOut™ Serum Replacement (KSR) (Cat. No. 10828-028)
- MEM Non-Essential Amino Acids Solution, 10 mM (Cat. No. 11140-050)
- Basic Fibroblast Growth Factor (bFGF) (Cat. No. PHG0264)
- β-mercaptoethanol, 1000X (Cat. No. 21985-023)

Continued on next page

Required materials, continued

- *Optional*: Penicillin-Streptomycin, Liquid (Cat. No. 15140-122)
- Polybrene Hexadimethrine Bromide (Sigma Cat. No. H9268)
- Attachment Factor (Cat. No. S006100)
- TrypLE™ Select Cell Dissociation Reagent (Cat. No. 12563) or 0.05% Trypsin/EDTA (Cat. No. 25300)
- Dulbecco's PBS (DPBS) without Calcium and Magnesium (Cat. No. 14190)

Reprogram StemPro™ CD34+ cells

The following protocol has been optimized for StemPro™ CD34+ cells derived from the human umbilical cord blood of mixed donors. Note that experimental conditions may vary among target cells and need to be optimized for each cell type. The example given in the following protocol does not guarantee the generation of iPSCs for all cell types.

Day –3: Seed cells

1. Three days before transduction, remove one vial of StemPro™ CD34+ cells (0.5×10^6 cells) from the liquid nitrogen storage tank.
2. Briefly roll the cryovial between hands to remove frost, and swirl it gently in a 37°C water bath to thaw the StemPro™ CD34+ cells.
3. When only a small ice crystal remains in the vial, remove it from water bath. Spray the outside of the vial with 70% ethanol before placing it in the cell culture hood.
4. Pipet the thawed cells gently into a 15-mL conical tube.
5. Add 10 mL of pre-warmed complete StemPro™-34 medium (see page 54) **drop-wise** to the cells. Gently mix by pipetting up and down.
   Note: Adding the medium slowly helps the cells to avoid osmotic shock.
6. Centrifuge the cell suspension at 200 × *g* for 10 minutes.
7. Discard the supernatant and resuspend the cells in 1 mL of complete StemPro™-34 medium containing cytokines (i.e., SCF, IL-3 and GM-CSF) (see page 54).
8. Place 0.5 mL each of cell suspension into two wells of a 24-well plate and incubate at 37°C in a humidified atmosphere of 5% CO_2.
   Note: We recommend using the wells in the middle section of the 24-well plate to prevent excessive evaporation of the medium during incubation.

Day –2: Observe cells and add fresh medium

9. Two days before transduction, add 0.5 mL of fresh complete StemPro™-34 medium containing cytokines without disturbing the cells. If cells are present in 0.5 mL removed from the wells, centrifuge the cell suspension at 200 × g for 10 minutes, discard the supernatant, and resuspend the cells in 0.5 mL fresh PBMC medium before adding them back to the plate.

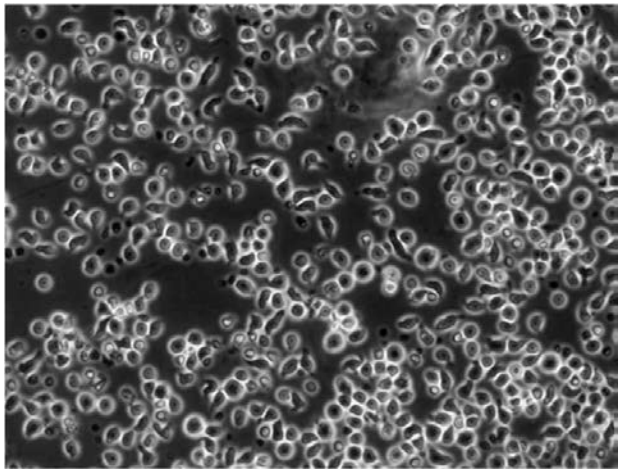
Continued on next page

Reprogram StemPro™ CD34+ cells

Day –1: Observe cells and add fresh medium

10. One day before transduction, gently remove 0.5 mL of medium and add 1 mL of fresh complete StemPro™-34 medium containing cytokines without disturbing the cells.

Figure 3 CD34+ cells grown in StemPro™-34 SFM.



Day 0: Count cells and perform transduction

11. Count the cells using the desired method (e.g., Countess™ Automated Cell Counter), and calculate the volume of each virus needed to reach the target MOI using the **live cell count** and the titer information on the CoA.

$$\text{Volume of virus (µL)} = \frac{\text{MOI (CIU/cell)} \times \text{number of cells}}{\text{titer of virus (CIU/mL)} \times 10^{-3}\ \text{(mL/µL)}}$$

Note: We recommend initially performing the transductions with MOIs of 5, 5, and 3 (i.e., KOS MOI=5, hc-Myc MOI=5, hKlf4 MOI=3). These MOIs can be optimized for your application.

Note: The titer of each CytoTune™ 2.0 reprogramming vector is lot-dependent. For the specific titer of your vectors, go to **thermofisher.com/cytotune** and search for the CoA by product lot number, which is printed on the vial. Avoid re-freezing and thawing of the reprogramming vectors since viral titers can decrease dramatically with each freeze/thaw cycle.

12. Harvest the cells and seed the necessary number of wells of a 24-well plate in a minimal volume (~100 µL) with 1.0×10^5 cells/well for transduction.
13. Remove one set of CytoTune™ 2.0 Sendai tubes from the –80°C storage. Thaw each tube one at a time by first immersing the bottom of the tube in a 37°C water bath for 5–10 seconds, and then removing the tube from the water bath and allowing it to thaw at room temperature. Once thawed, briefly centrifuge the tube and place it immediately on ice.

Continued on next page

Reprogram StemPro™ CD34+ cells, continued

14. Add the calculated volumes of each of the three CytoTune™ 2.0 Sendai viruses to 0.4 mL of pre-warmed StemPro™-34 medium containing cytokines and 4 µg/mL of Polybrene. Ensure that the solution is thoroughly mixed by pipetting the mixture gently up and down. Complete the next step within 5 minutes.
15. Add the reprogramming virus mixture (from Step 14) to the well(s) containing cells (from Step 12). Incubate the cells at 37°C in a humidified atmosphere of 5% CO_2 overnight.

Day 1: Replace medium and culture cells

16. Remove the CytoTune™ 2.0 Sendai viruses by centrifuging the cells at 400 × *g* for 10 minutes. Aspirate and discard the supernatant.
17. Resuspend the cells in 0.5 mL of complete StemPro™-34 Medium containing cytokines (see page 54) in the 24-well plate.
18. Incubate the cells in at 37°C in a humidified atmosphere of 5% CO_2 for two days.

    Note: While the cells are incubating (i.e., 1–2 days before passaging the transduced cells), prepare the necessary number of MEF culture dishes for each well containing transduced cells (see page 55).

Day 3: Plate cells on MEF dishes

19. Count the cells using the desired method (e.g., Countess™ Automated Cell Counter) and seed the MEF dishes with at least two different densities between 1 x 10^4 –1 X 10^5 $CD34^+$ cells per well of a 6-well in 2 mL of complete StemPro™-34 Medium without cytokines.

    Note: It is strongly recommended to set aside cells at this point for RNA extraction to be used as a positive control in the RT-PCR or qPCR detection of the CytoTune™ vectors. It is very important to include this positive control when performing detection of the CytoTune™ vectors.
20. Incubate the cells at 37°C in a humidified atmosphere of 5% CO_2 for three days.
21. Replace half of the spent medium every other day. Gently remove 1 mL of medium from the cells and replace with 1 mL of complete StemPro™-34 Medium without cytokines.

    Note: The transduced cells may only be loosely attached for the first few days after plating. Be sure to perform media changes gently during this time. If cells are present in 1 mL removed from the wells, centrifuge the cell suspension at 200 × g for 10 minutes, discard the supernatant, and resuspend the cells in 1 mL fresh medium before adding them back to the plate.

Day 7: Transition to iPSC medium

22. Remove 1 mL of medium from the cells and add 1 mL of iPSC medium (see page 53) to transition the cells to the new culture medium.
23. Incubate the cells in a 37°C, 5% CO_2 incubator overnight.

Continued on next page

Reprogram StemPro™ CD34+ cells, continued

Day 8 to 28: Feed and Monitor the Cells

24. The next day, remove the spent medium completely and replace with 2 mL of iPSC medium. Replace spent medium daily.

    Note: Plated cells should have fully attached by this point. Any cells that have not attached will likely never do so.

25. Starting on Day 8, observe the plates every other day under a microscope for the emergence of cell clumps indicative of reprogrammed cells (see Figure 4, below).

    Note: For BJ fibroblasts (positive control), colony formation is normally observed on Day 12 post-transduction. However, depending on cell type, it may take up to 4 weeks before colonies are seen.

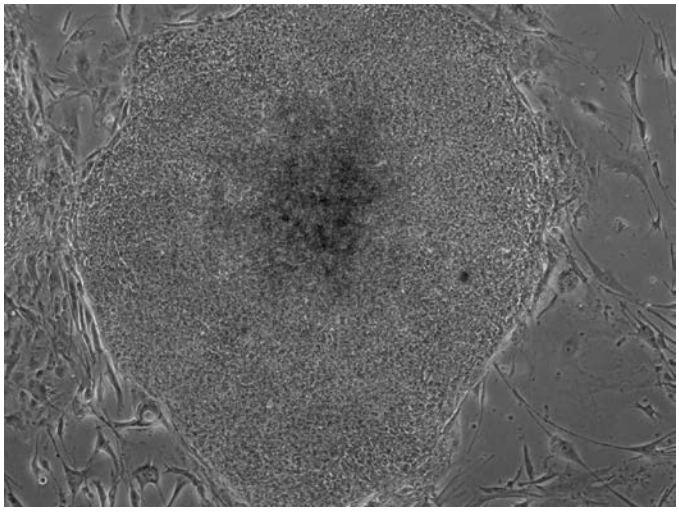
26. Day 15 to 20 after transduction, colonies should have grown to an appropriate size for transfer. The day before transferring the colonies, prepare the necessary number of MEF plates using Attachment Factor-coated 12- or 6-well plates.

    Note: We recommend harvesting colonies closer to 3 weeks to avoid differentiation.

27. When colonies are ready for transfer, perform live staining using Tra1-60 or Tra1-81 for selecting reprogrammed colonies if desired (see **Live stain**, page 46).

28. Manually pick colonies and transfer them onto prepared MEF dishes (see **Pick iPSC colonies**, page 47).

Figure 4 Emergence of cell clumps indicative of reprogrammed cells at Day 19.

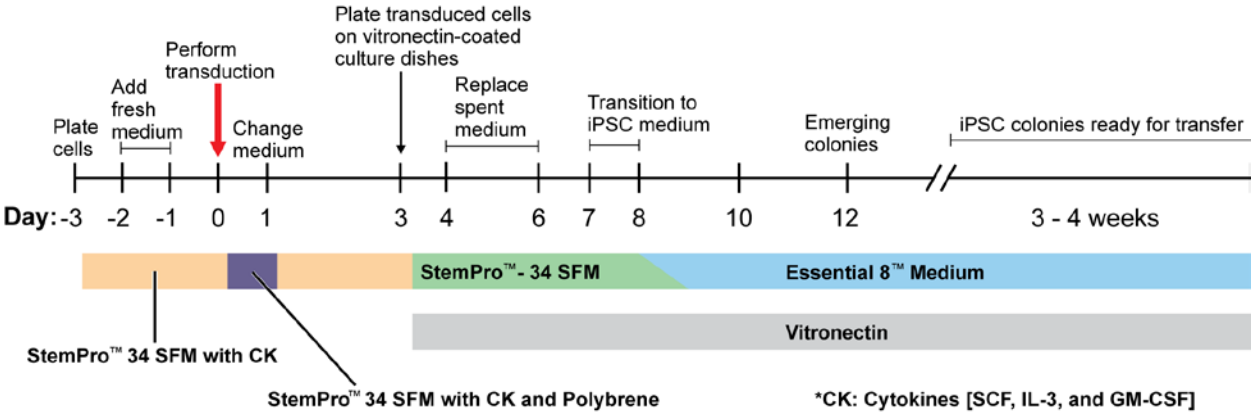


Experiment outline (Feeder-Free)

Workflow

The major steps required for reprogramming StemPro™ CD34+ cells using the CytoTune™-iPS 2.0 Sendai Reprogramming Kit to generate iPSCs cultured on vitronectin-coated culture dishes are shown below.

Note: The timeline is provided as a guideline for experimental planning; actual timeline can vary based on the cell type and experimental conditions.



Reprogramming timeline

Day –3: Plate StemPro™ CD34+ cells into at least two wells of a 24-well plate in complete StemPro™-34 medium containing cytokines (i.e., SCF, IL-3 and GM-CSF).

Day –1: Replace 0.5 mL of spent medium with 0.5 mL of fresh complete StemPro™-34 medium containing cytokines.

Day –2: Replace 0.5 mL of spent medium with 1 mL of fresh complete StemPro™-34 medium containing cytokines.

Day 0: Transduce the cells using the CytoTune™ 2.0 Sendai reprogramming vectors at the appropriate MOI in StemPro™-34 medium containing cytokines and 4 µg/mL of Polybrene. Incubate the cells overnight.

Day 1: Replace the medium with fresh StemPro™-34 medium containing cytokines (no Polybrene) to remove the CytoTune™ 2.0 Sendai reprogramming vectors.

Day 3: Plate the transduced cells on vitronectin-coated culture dishes in complete StemPro™-34 medium without cytokines.

Day 4–6: Replace half of the spent medium with fresh complete StemPro™-34 medium without cytokines every other day.

Day 7: Start transitioning into Essential 8™ Medium by replacing half of the StemPro™-34 medium without cytokines with complete iPSC medium.

Day 8: Replace the entire medium with Essential 8™ Medium to conclude the transitioning and continue culturing cells on vitronectin-coated culture dishes.

Day 9–28: Replace spent medium with fresh Essential 8™ Medium every day and monitor the culture vessels for the emergence of iPSC colonies. When iPSC colonies are ready for transfer, perform live staining, and pick and transfer undifferentiated iPSCs onto fresh vitronectin-coated culture dishes for expansion.

Reprogram StemPro™ CD34+ Cells (Feeder-Free)

Media for reprogramming StemPro™ CD34+ cells (feeder-free)

For optimal reprogramming of CD34+ cells using the CytoTune™-iPS 2.0 Sendai Reprogramming Kit to generate iPSCs cultured on vitronectin-coated plates, use the following media at the designated stages of the reprogramming experiment:

- **StemPro™-34 medium containing cytokines (i.e., SCF, IL-3 and GM-CSF)** (page 54): Plating cells prior to transduction, post-transduction recovery of cells
- **StemPro™-34 medium containing cytokines + 4 µg/mL of Polybrene**: Transduction
- **StemPro™-34 medium without cytokines** (page 54): Plating of transduced cells on MEF culture dishes
- **Complete Essential 8™ Medium** (page 53): Expansion of transduced cells on vitronectin-coated culture dishes, live staining and picking of iPSCs

Required materials

Cells and vectors

- CytoTune™ 2.0 Sendai reprogramming vectors
  Note: For successful reprogramming, you need all three tubes of reprogramming vectors.
- StemPro™ CD34+ cells to reprogram
  Note: StemPro™ CD34+ cells are available as part of the StemPro™-34 Medium and CD34+ Cell Kit (Cat. No. A14059).
- *Optional*: Human neonatal foreskin fibroblast cells (strain BJ; ATCC no. CRL2522) as a positive reprogramming control
  Note: If you are using this as a control, follow the protocol for reprogramming fibroblasts within this manual (page 12).

Media and reagents

- StemPro™-34 Medium and CD34+ Cell Kit (Cat. No. A14059)
- Recombinant Human SCF Lyophilized (Cat. No. PHC2111)
- Recombinant Human IL-3 Lyophilized (Cat. No. PHC0031)
- Recombinant Human GM-CSF Lyophilized (Cat. No. PHC2011)

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Required materials, continued

- *Optional*: Penicillin-Streptomycin, Liquid (Cat. No. 15140-122)
- Polybrene Hexadimethrine Bromide (Sigma Cat. No. H9268)
- Dulbecco's PBS (DPBS) without Calcium and Magnesium (Cat. No. 14190)

Reprogram StemPro™ CD34+ cells

The following protocol has been optimized for StemPro™ CD34+ cells derived from the human umbilical cord blood of mixed donors. Note that experimental conditions may vary among target cells and need to be optimized for each cell type. The example given in the following protocol does not guarantee the generation of iPSCs for all cell types.

Day –3: Seed cells

1. Three days before transduction, remove one vial of StemPro™ CD34+ cells (0.5×10^6 cells) from the liquid nitrogen storage tank.
2. Briefly roll the cryovial between hands to remove frost, and swirl it gently in a 37°C water bath to thaw the StemPro™ CD34+ cells.
3. When only a small ice crystal remains in the vial, remove it from water bath. Spray the outside of the vial with 70% ethanol before placing it in the cell culture hood.
4. Pipet the thawed cells gently into a 15-mL conical tube.
5. Add 10 mL of pre-warmed complete StemPro™-34 medium (see page 54) **drop-wise** to the cells. Gently mix by pipetting up and down.
   Note: Adding the medium slowly helps the cells to avoid osmotic shock.
6. Centrifuge the cell suspension at 200 × *g* for 10 minutes.
7. Discard the supernatant and resuspend the cells in 1 mL of complete StemPro™-34 medium containing cytokines (i.e., SCF, IL-3 and GM-CSF) (see page 54).
8. Place 0.5 mL each of cell suspension into two wells of a 24-well plate and incubate at 37°C in a humidified atmosphere of 5% CO_2.
   Note: We recommend using the wells in the middle section of the 24-well plate to prevent excessive evaporation of the medium during incubation.

Day –2: Observe cells and add fresh medium

9. Two days before transduction, add 0.5 mL of fresh complete StemPro™-34 medium containing cytokines without disturbing the cells. If cells are present in 0.5 mL removed from the wells, centrifuge the cell suspension at 200 × g for 10 minutes, discard the supernatant, and resuspend the cells in 0.5 mL fresh PBMC medium before adding them back to the plate.

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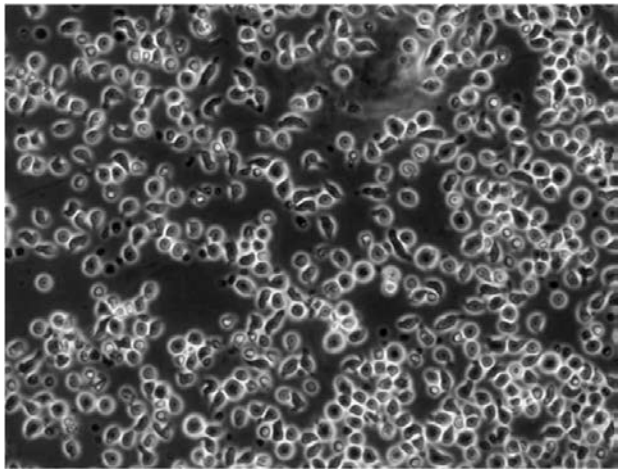
Reprogram StemPro™ CD34+ cells

Day –1: Observe cells and add fresh medium

10. One day before transduction, gently remove 0.5 mL of medium and add 1 mL of fresh complete StemPro™-34 medium containing cytokines without disturbing the cells.

Figure 3 CD34+ cells grown in StemPro™-34 SFM.



Day 0: Count cells and perform transduction

11. Count the cells using the desired method (e.g., Countess™ Automated Cell Counter), and calculate the volume of each virus needed to reach the target MOI using the **live cell count** and the titer information on the CoA.

$$\text{Volume of virus (µL)} = \frac{\text{MOI (CIU/cell)} \times \text{number of cells}}{\text{titer of virus (CIU/mL)} \times 10^{-3}\ \text{(mL/µL)}}$$

Note: We recommend initially performing the transductions with MOIs of 5, 5, and 3 (i.e., KOS MOI=5, hc-Myc MOI=5, hKlf4 MOI=3). These MOIs can be optimized for your application.

Note: The titer of each CytoTune™ 2.0 reprogramming vector is lot-dependent. For the specific titer of your vectors, go to **thermofisher.com/cytotune** and search for the CoA by product lot number, which is printed on the vial. Avoid re-freezing and thawing of the reprogramming vectors since viral titers can decrease dramatically with each freeze/thaw cycle.

12. Harvest the cells and seed the necessary number of wells of a 24-well plate in a minimal volume (~100 µL) with 1.0×10^5 cells/well for transduction.
13. Remove one set of CytoTune™ 2.0 Sendai tubes from the –80°C storage. Thaw each tube one at a time by first immersing the bottom of the tube in a 37°C water bath for 5–10 seconds, and then removing the tube from the water bath and allowing it to thaw at room temperature. Once thawed, briefly centrifuge the tube and place it immediately on ice.

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Reprogram StemPro™ CD34+ cells, continued

14. Add the calculated volumes of each of the three CytoTune™ 2.0 Sendai viruses to 0.4 mL of pre-warmed StemPro™-34 medium containing cytokines and 4 µg/mL of Polybrene. Ensure that the solution is thoroughly mixed by pipetting the mixture gently up and down. Complete the next step within 5 minutes.
15. Add the reprogramming virus mixture (from Step 14) to the well(s) containing cells (from Step 12). Incubate the cells at 37°C in a humidified atmosphere of 5% CO_2 overnight.

Day 1: Replace medium and culture cells

16. Remove the CytoTune™ 2.0 Sendai viruses by centrifuging the cells at 400 × *g* for 10 minutes. Aspirate and discard the supernatant.
17. Resuspend the cells in 0.5 mL of complete StemPro™-34 Medium containing cytokines (see page 54) in the 24-well plate.
18. Incubate the cells in at 37°C in a humidified atmosphere of 5% CO_2 for two days.

Day 3: Plate cells on MEF dishes

19. Coat a sufficient number of tissue culture dishes (e.g. 6-well, 60-mm, or 100-mm) with vitronectin (see page 57 for coating protocol).

    Note: Geltrex™ Membrane Matrix can be substituted for vitronectin (see page 59 for coating protocol).
20. Count the cells using the desired method (e.g., Countess™ Automated Cell Counter) and seed the vitronectin-coated dishes with at least two different densities between 1 x 10^4 –1 X 10^5 $CD34^+$ cells per well of a 6-well in 2 mL of complete StemPro™-34 Medium without cytokines.

    Note: It is strongly recommended to set aside cells at this point for RNA extraction to be used as a positive control in the RT-PCR or qPCR detection of the CytoTune™ vectors). It is very important to include this positive control when performing detection of the CytoTune™ vectors.
21. Incubate the cells at 37°C in a humidified atmosphere of 5% CO_2 for three days.
22. Replace half of the spent medium every other day. Gently remove 1 mL of medium from the cells and replace with 1 mL of complete StemPro™-34 Medium without cytokines.

    Note: The transduced cells may only be loosely attached for the first few days after plating. Be sure to perform media changes gently during this time. If cells are present in 1 mL removed from the wells, centrifuge the cell suspension at 200 × g for 10 minutes, discard the supernatant, and resuspend the cells in 1 mL fresh medium before adding them back to the plate.

Day 7: Transition to Essential 8™ Medium

23. Remove 1 mL of medium from the cells and add 1 mL of Essential 8™ Medium (see page 53) to transition the cells to the new culture medium.
24. Incubate the cells in a 37°C, 5% CO_2 incubator overnight.

Continued on next page

Reprogram StemPro™ CD34+ cells, continued

Day 8 to 28: Feed and Monitor the Cells

25. The next day, remove the spent medium completely and replace with 2 mL of Essential 8™ Medium. Replace spent medium daily.

    Note: Plated cells should have fully attached by this point. Any cells that have not attached will likely never do so.

26. Starting on Day 8, observe the plates every other day under a microscope for the emergence of cell clumps indicative of reprogrammed cells.

    Note: For BJ fibroblasts (positive control), colony formation is normally observed on Day 12 post-transduction. However, depending on cell type, it may take up to 4 weeks before colonies are seen.

27. Day 15 to 20 after transduction, colonies should have grown to an appropriate size for transfer.

    Note: We recommend harvesting colonies closer to 3 weeks to avoid differentiation.

28. When colonies are ready for transfer, perform live staining using Tra1-60 or Tra1-81 for selecting reprogrammed colonies if desired (see **Live stain**, page 46).

29. Manually pick colonies and transfer them onto prepared vitronectin-coated 12- or 6-well culture dishes (see **Pick iPSC colonies**, page 47).

Visual identification

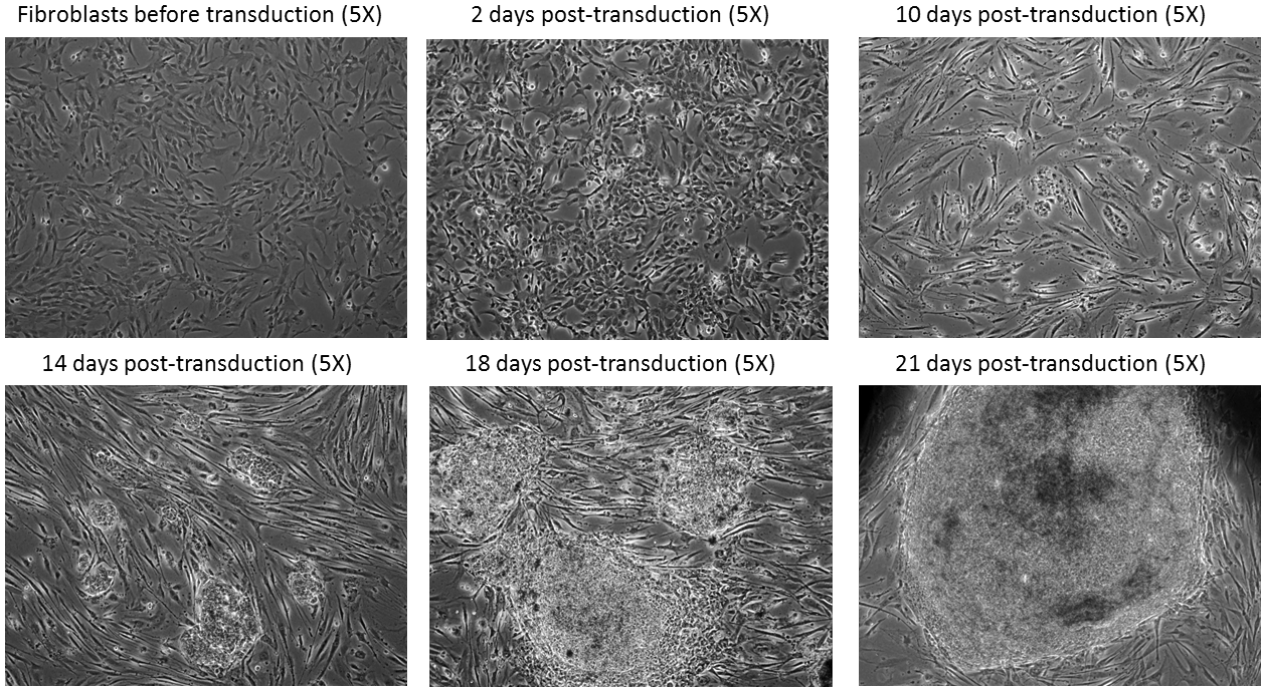
Visual identification

By Day 21 post-transduction, the cell colonies on the MEF culture dishes will have become large and compact, covering the majority of the surface area of the culture dish. However, only a fraction of these colonies will consist of iPSCs, which exhibit a hESC-like morphology characterized by a flatter cobblestone-like appearance with individual cells clearly demarcated from each other in the colonies (see Figure 5, below). Therefore, we recommend that you perform live staining with Tra1-60 or Tra1-81 antibodies that recognize undifferentiated hESCs (see **Live stain**, page 46).

Morphology of reprogrammed cells

The images below show the morphology of human neonatal foreskin fibroblast cells (strain BJ) that were reprogrammed into iPSCs using the CytoTune™-iPS 2.0 Sendai Reprogramming Kit.

Figure 5 Human neonatal foreskin fibroblast cells (strain BJ) were transduced using the CytoTune™-iPS 2.0 Sendai Reprogramming Kit and allowed to proliferate on MEF feeder layers in iPSC medium. The images were obtained using a 5X objective.



Live stain

Live stain with antibodies

One of the fastest and most reliable methods for identifying a reprogrammed colony is live staining with Tra1-60 or Tra1-81 antibodies that recognize undifferentiated iPSCs and enable the identification of reprogrammed cells from a variety of human cell types.

Note: Other methods of identifying iPSCs (such as alkaline phosphatase staining) are also acceptable.

Required antibodies

TRA-1-60 Mouse anti-human mAb, AlexaFluor™ 488 Conjugate Kit for Live Cell Imaging (Cat. No. A25618)

or

TRA-1-60 Mouse anti-human mAb, Alexa Fluor™ 594 Conjugate Kit for Live Cell Imaging (Cat. No.A24882)

Live stain cells

If live-stained cells are to be used for further culture, be sure to use antibodies that are sterile (filter sterilize as necessary) and work aseptically. .

1. Centrifuge the dye-conjugated antibody solution (e.g., 2 minutes at 10,000 × g) and only use the supernatant.

   Note: This step minimizes transferring protein aggregates that may have formed during storage, thereby reducing non-specific background staining.

2. Add a 1:50 volume of the dye-conjugated antibody directly to the cell culture medium of the cells to be stained (see Table I), then mix by gentle swirling.
3. Incubate for 30 minutes at 37°C.
4. Remove the staining solution and gently wash the cells 2–3 times with FluoroBrite™ DMEM (Cat. No A1896701).
5. For optimal results, image the cells immediately (i.e., within 30 minutes).

   Note: To continue culturing the cells, replace the FluoroBrite™ DMEM with fresh cell culture medium and return the cells to the 37°C incubator.

Table I Recommended volumes for staining protocol

Culture format	No. of tests	Staining volume	50X antibody
96-well plate	200	50 µL/well	1 µL
48- well plate	100	100 µL/well	2 µL
24- well plate	50	200 µL/well	4 µL
12- well plate	25	400 µL/well	8 µL
6- well plate	10	1 mL/well	20 µL
35-mm dish	10	1 mL/dish	20 µL
60-mm dish	5	2 mL/dish	40 µL
100-mm dish	2	5 mL/dish	100 µL
4-well chamber slide	25	400 µL/well	8 µL
8-well chamber slide	50	200 µL/well	4 µL

Pick iPSC colonies

Pick iPSC colonies (feeder-dependent)

1. Place the culture dish containing the reprogrammed cells under an inverted microscope and examine the colonies under 10X magnification.
2. Mark the colony to be picked on the bottom of the culture dish.

   Note: We recommend picking at least 10 distinct colonies by the end of each reprogramming experiment and expanding them in separate 12- or 6-well MEF culture plates (see below).
3. Transfer the culture dish to a sterile cell culture hood (i.e., biosafety cabinet) equipped with a stereomicroscope.
4. Using a 25 gauge 1½ inch needle, cut the colony to be picked into 5–6 pieces in a grid-like pattern.
5. Using a 200 µL pipette, transfer the cut pieces to a freshly prepared 12- or 6-well MEF culture plate (see page 55) containing iPSC medium (see page 53).
6. Incubate the MEF culture plate containing the picked colonies in a 37°C incubator with a humidified atmosphere of 5% CO_2.
7. Allow the colonies to attach to the culture plate for 48 hours before replacing the spent medium with fresh iPSC medium. After that, change the medium every day.
8. Treat the reprogrammed colonies like normal human ESC colonies and passage, expand, and maintain them using standard culture procedures until you have frozen cells from two 60-mm plates (see **Freezing iPSCs in iPSC freezing medium**, page 64).

Pick iPSC colonies (feeder-free)

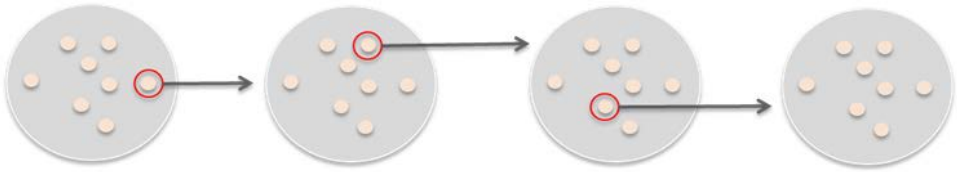
1. Pick the iPSCs as described on above, up to step 4.
2. Using a 200 µL pipette, transfer the cut pieces onto a vitronectin-coated 12- or 6-well culture plate (see page 57) containing complete Essential 8™ Medium (page 53).
3. Incubate the vitronectin-culture plate containing the picked colonies in a 37°C incubator with a humidified atmosphere of 5% CO_2.
4. Allow the colonies to attach to the culture plate for 48 hours before replacing the spent medium with fresh complete Essential 8™ Medium. After that, change the medium every day.
5. When the colonies cover ~85% of the surface area of the culture vessel, they are ready for passaging. Passage the colonies using 0.5 mM EDTA prepared in Dulbecco's Phosphate-Buffered Saline (DPBS) without calcium or magnesium (see **Passage iPSCs with EDTA**, page 63).

   Note: Enzymes such as collagenase and dispase do not work well with cells cultured in Essential 8™ Medium on vitronectin-coated plates. Use of these enzymes for passaging cells results in compromised viability and attachment.
6. Continue to culture, expand, and maintain the reporgrammed colonies in complete Essential 8™ Medium until you have frozen cells from two 60-mm plates (see **Freeze iPSCs in Essential 8™ freezing medium**, page 65).

Generate vector-free iPSCs

Guidelines for generating vector-free iPSCs

- The time needed to derive vector-free iPSCs may vary depending on culture and passage conditions. In the case of human neonatal foreskin fibroblast cells (strain BJ), it takes about 1–2 months after gene transduction to obtain iPSCs free of CytoTune™ 2.0 Sendai reprogramming vectors.
- To obtain virus-free clones faster, we recommend that you perform single colony subcloning for the first few passages (minimum 5) instead of bulk or pooled-clone passaging.
- To perform single colony subcloning, pick from a single colony to transfer to another 6-well plate (Passage 1). From Passage 1, pick a single colony and transfer to another 6-well plate (passage 2) and so forth. We recommend subcloning for 5 passages and then testing for virus free iPSCs.



Required materials

- Dulbecco's PBS (DPBS) without Calcium and Magnesium (Cat. No. 14190)
- Rabbit anti-SeV antibody (MBL International Corporation; Cat. no PD029)
- Alexa Fluor™ 488 goat anti-rabbit IgG (H+L) antibody (Cat no. A11034) or Alexa Fluor™ 594 goat anti-rabbit IgG (H+L) antibody (Cat no. A11037)
- TRIzol™ LS reagent (Cat. No. 10296-010)
- SuperScript™ VILO™ cDNA Synthesis Kit (Cat. No. 11754-050)
- AccuPrime™ SuperMix I (Cat. No. 12342-010)

Generate vector-free iPSCs

1. When passaging iPSC colonies, prepare duplicate plates; one for immunostaining and one for further passaging.
2. Perform immunostaining on one plate using anti-SeV antibodies (see below).
3. If any colonies stain positive, perform cell cloning on the other duplicate plate.
4. Repeat immunostaining with anti-SeV antibodies on the cloned colonies until all colonies in a plate are negative.
5. If all colonies are negative for anti-SeV antibodies, passage the cells and confirm the absence of the CytoTune™ 2.0 Sendai reprogramming vectors by RT-PCR (see page 49), or real-time RT-PCR.

Continued on next page

Immunocyto-chemistry with anti-SeV antibodies

1. Wash cells once with D-PBS
2. Fix the cells in 4% paraformaldehyde for 5 minutes at room temperature.
3. Wash cells twice with D-PBS.
4. Add the anti-SeV antibody (MBL, Cat. no PD029) diluted in 0.1% Triton™ X-100 in D-PBS to the cells and incubate for 1 hour at 37°C.
5. Remove the antibody solution. Wash the cells 3 times with D-PBS.
6. Add the secondary antibody diluted in 0.1% Triton™ X-100 in D-PBS to the cells and incubate for 1 hour at 37°C.
7. Remove the secondary antibody solution from the dish. Wash the cells 3 times with D-PBS.
8. Visualize the cells under a fluorescence microscope.

RT-PCR protocol for detecting the SeV genome and transgenes

1. Extract the total RNA from 5×10^6 iPSCs using the TRIzol™ Reagent (Cat. No. 15596-026) following the instructions provided with the reagent. As a positive control, use cells set aside during the reprogramming procedure.

   Note: It is important to use cells set aside during the reprogramming experiment as a valid positive control. Early-passage iPSC may already have cleared one or more of the vectors and are therefore not a suitable positive control.
2. Carry out a reverse transcription reaction using 1 µg of RNA (from step 1) and the SuperScript™ VILO™ cDNA Synthesis Kit (Cat. No. 11754-050) following the instructions provided with the kit.

   Note: Because the CytoTune™ 2.0 Sendai reprogramming vectors are based on SeV, which is an RNA virus, reverse transcription is required for detecting the presence of the SeV genome in your reprogrammed cells.
3. Carry out the PCR using 10 µL of cDNA from the reverse transcription reaction (Step 2, above) and AccuPrime™ SuperMix I (Cat. No. 12342-010) with the parameters below. For the RT-PCR primer sequences and the expected product size, refer to the table on page 50.

Step	Temperature	Time	Cycles
Denaturation	95°C	30 seconds	30–35
Annealing	55°C	30 seconds	
Elongation	72°C	30 seconds	

4. Analyze the PCR products using 2% agarose gel electrophoresis.

If you still detect CytoTune™ 2.0 Sendai virus in your iPSC lines after more than 10 passages, and have performed RT-PCR to show that hKlf4 is absent from your cells (this vector does not have the temperature sensitive mutation), then you can perform a temperature shift to remove KOS and/or the c-Myc vector(s). CytoTune™ 2.0 Sendai hc-Myc tends to persist in the cells longer than the other CytoTune™ 2.0 Sendai reprogramming vectors. However, because this vector contains a temperature sensitivity mutation, you can enhance its removal and obtain complete absence of Sendai virus by incubating your cells at 38–39°C for 5 days.

RT-PCR primer sets

Use the following RT-PCR primer sets to detect the SeV genome and transgenes in cells reprogrammed using the CytoTune™ 2.0 Sendai reprogramming vectors.

Target	Primer sets	Product size
SeV	Forward: GGA TCA CTA GGT GAT ATC GAG C* Reverse: ACC AGA CAA GAG TTT AAG AGA TAT GTA TC*	181 bp
KOS	Forward: ATG CAC CGC TAC GAC GTG AGC GC Reverse: ACC TTG ACA ATC CTG ATG TGG	528 bp
Klf4	Forward: TTC CTG CAT GCC AGA GGA GCC C Reverse: AAT GTA TCG AAG GTG CTC AA*	410 bp
c-Myc	Forward: TAA CTG ACT AGC AGG CTT GTC G* Reverse: TCC ACA TAC AGT CCT GGA TGA TGA TG	532 bp

* Primer contains SeV genome sequences. Pairing of these primers with transgene-specific primers allows specific detection of transgenes carried by the CytoTune™ 2.0 Sendai reprogramming vectors.

Real time RT-PCR

As an alternative to endpoint RT-PCR, use the following pre-validated TaqMan™ assays to detect the presence of CytoTune™ 2.0 Sendai reprogramming vectors by real-time RT-PCR:

Target	Taqman assay
SeV	Mr04269880_mr
KOS	Mr04421257_mr
Klf4	Mr04421256_mr
cMyc	Mr04269876_mr

Problem	Possible cause	Solution
Cytotoxic effects observed after transduction	Viral load too high	Decrease the volume of CytoTune™ 2.0 vector or increase the starting cell number.
Too many colonies on the plate	Too many cells plated	Decrease the number of cells plated after transduction.
No iPSC colony formation	Insufficient amount of virus used	• Check the volume of the CytoTune™ 2.0 vector and the starting cell number. Changing the MOI may improve the results. • We suggest initially increasing the MOI of Klf4 to improve efficiency (e.g., KOS MOI = 5, c-Myc MOI =5, Klf4 MOI =6). If efficiencies are still too low, increase the MOI of KOS and c-Myc, while maintaining a 1:1 ratio between the two (e.g. KOS MOI = 10, c-Myc MOI = 10, Klf4 =6). • Not all cell types will be reprogrammed with the same efficiency. Check the levels of protein expression in your cell type using TaqMan™ Protein Assays (see page 68 for ordering information). • Do not re-freeze thaw or aliquot virus. Viral titer is not guaranteed for kits refrozen or thawed.
Too few iPSC colony compared to BJ fibroblasts	Cell type not efficiently reprogrammable	• Not all cell types will have the same reprogramming efficiency. Increase the number of the cells plated. • If reprogramming a new cell type, use the CytoTune™-EmGFP Sendai Fluorescence Reporter to assess uptake of Sendai virus (page 60).
iPSC colonies look differentiated	iPSC colonies transferred to MEF dishes too late	Perform staining earlier and transfer iPSC colony to fresh feeder cells.
Difficult to obtain vector-free iPSCs	Cell type cannot efficiently eliminate the CytoTune™ 2.0 Sendai reprogramming vector	• Some cell strains may need longer time to eliminate the CytoTune™ 2.0 Sendai vectors and become vector-free compared to other strains. Perform repeated single colony subcloning until you obtain negative cells as determined by immunocytochemistry with anti-SeV antibodies. • It may be easier to obtain SeV-negative colonies if single colony subcloning is performed by transferring a portion of a colony with a glass pipette. • The rate with which iPSC colonies eliminate the CytoTune™ 2.0 Sendai vectors may increase if the cells are incubated for 5 days at 38–39°C **after** you have confirmed by RT PCR that the Klf4 vector is absent from your cells and only KOS and/or c-Myc vectors remain.

Prepare media and reagents

Basic FGF stock solution

1. To prepare 1000 µL of 10-µg/mL Basic FGF solution, aseptically mix the following components:

Basic FGF	10 µg
DPBS without Calcium and Magnesium	980 µL
10% BSA	10 µL

2. Aliquot and store the Basic FGF solution at –20°C for up to 6 months.

SCF (c-kit Ligand), FLT-3 Ligand, IL-3, IL-6, and GM-CSF Stock Solutions

SCF (c-kit Ligand), FLT-3 Ligand, IL-3, IL-6, and GM-CSF are supplied lyophilized. Prepare stock solutions as described in their specific product inserts and store small aliquots frozen. Thaw at time of use.

Collagenase IV solution

1. To prepare 50 mL of a 1-mg/mL Collagenase IV solution, aseptically mix the following components:

Collagenase IV	50 mg
DMEM/F-12	50 mL

2. Sterilize the Collagenase IV solution through a 0.2 µm filter.
3. Aliquot and store the Collagenase IV solution at –20°C for up to 6 months.

0.5 mM EDTA in DPBS (50 mL)

1. To prepare 50 mL of 0.5 mM EDTA in DPBS, aseptically mix the following components in a 50-mL conical tube in a biological safety cabinet:

DPBS without Calcium and Magnesium	50 mL
0.5 M EDTA	50 µL

2. Filter sterilize the solution. The solution can be stored at room temperature for up to six months.

MEF/Fibroblast medium (for 100 mL of complete medium)

3. To prepare 100 mL of complete MEF/fibroblast medium, aseptically mix the following components:

DMEM	89 mL
FBS, ESC-Qualified	10 mL
MEM Non-Essential Amino Acids Solution, 10 mM	1 mL
β-mercaptoethanol, 55 mM	100 µL

4. Complete MEF/fibroblast medium can be stored at 2–8°C for up to 1 week.

Continued on next page

iPSC medium (for 100 mL of complete medium)

1. To prepare 100 mL of complete iPSC medium, aseptically mix the following components:

DMEM/F-12, GlutaMAX™ supplement	78 mL
KnockOut™ Serum Replacement (KSR)	20 mL
MEM Non-Essential Amino Acids Solution, 10 mM	1 mL
β-mercaptoethanol, 55 mM	100 µL
Penicillin-Streptomycin (optional)	1 mL
bFGF (10 µg/mL)*	40 µL

   * Prepare the iPSC medium without bFGF, and then supplement with fresh bFGF when the medium is used.
2. Complete iPSC Medium (without bFGF) can be stored at 2–8°C for up to 1 week.

Essential 8™ Medium (for 500 mL of complete medium)

1. Thaw frozen Essential 8™ Supplement at 2–8°C overnight before using it to prepare complete medium. **Do not thaw the frozen supplement at 37°C.**
2. Mix the thawed supplement by gently inverting the vial a couple of times, remove 10 mL from the bottle of Essential 8™ Basal Medium, and then aseptically transfer the entire contents of the Essential 8™ Supplement to the bottle of Essential 8™ Basal Medium. Swirl the bottle to mix and to obtain 500 mL of homogenous complete medium.
3. Complete Essential 8™ Medium can be stored at 2–8°C for up to 2 weeks. Before use, warm complete medium required for that day at room temperature until it is no longer cool to the touch. **Do not warm the medium at 37°C.**

PBMC medium (for 500 mL of complete medium)

PBMC medium consists of complete StemPro™-34 medium supplemented with the appropriate cytokines. Follow the procedure below to prepare 500 mL of complete PBMC medium.

1. Thaw the frozen StemPro™-34 Nutrient Supplement at 4°C overnight.
2. After thawing, mix the supplement well by gently inverting the vial a couple of times, and then aseptically transfer the entire contents of the vial to the bottle of StemPro™-34 SFM. Swirl the bottle to mix and to obtain a homogenous complete medium.
3. Aseptically add L-Glutamine to a final concentration of 2 mM (5 mL of 200 mM L-Glutamine to 500 mL of medium).
4. The complete medium (without cytokines) has a shelf life of 30 days when stored at 2–8°C, in the dark.
5. Add the following cytokines to the indicated final concentration **on the day of use**:

Cytokine	**Final concentration**
SCF	100 ng/mL
FLT-3	100 ng/mL
IL-3	20 ng/mL
IL-6	20 ng/mL

StemPro™-34 medium (for 500 mL complete medium)

1. Thaw the frozen StemPro™-34 Nutrient Supplement overnight at 4°C.
2. Mix the thawed supplement well by gently inverting the vial several times. Aseptically transfer the entire contents of the vial to the bottle of StemPro™-34 SFM. Swirl the bottle to mix and to obtain a homogenous complete medium.
3. Aseptically add L-Glutamine to a final concentration of 2 mM (5 mL of 200 mM L-Glutamine to 500 mL of medium)

   Note: You may substitute L-Glutamine with 5 mL of GlutaMAX™-I (100X) in 500 mL of medium.
4. Complete StemPro™-34 medium (without cytokines) can be stored at 2–8°C for up to 4 weeks.

StemPro™-34 medium with cytokines for CD34+ cell culture (for 10 mL complete medium)

1. Prepare cytokines (SCF, IL-3, GM-CSF) according to their specific product instructions.
2. **On the day of use**, aseptically add the appropriate volume of each cytokine to StemPro™-34 Medium to achieve the recommended final concentration (10 mL total volume):

Cytokine	Final concentration
SCF	100 ng/mL
IL-3	50 ng/mL
GM-CSF	25 ng/mL

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iPSC freezing medium

Prepare the Freezing Media A and B immediately before use.

1. In a sterile 15-mL tube, mix together the following reagents for every 1 mL of **freezing medium A** needed:

iPSC medium	0.5 mL
KnockOut™ Serum Replacement	0.5 mL

2. In another sterile 15-mL tube, mix together the following reagents for every 1 mL of **freezing medium B** needed:

iPSC medium	0.8 mL
DMSO	0.2 mL

3. Place the tube with freezing medium B on ice until use (you can keep freezing medium A at room temperature). Discard any remaining freezing medium after use.

Essential 8™ freezing medium

1. Pre-warm the required volume of Essential 8™ Medium at room temperature until it is no longer cool to the touch. **Do not warm the medium in a 37°C water bath**.
2. Prepare Essential 8™ Freezing Medium. For every 1 mL of freezing medium needed, aseptically combine the components listed below in a sterile 15-mL tube:

Complete Essential 8™ Medium	0.9 mL
DMSO	0.1 mL

3. Place the tube with Essential 8™ Freezing Medium on ice until use. Discard any remaining freezing medium after use.

Prepare MEF culture dishes

Gelatin coating culture vessels

1. Cover the whole surface of each culture vessel with Attachment Factor (AF) solution and incubate the vessels for 30 minutes at 37°C or for 2 hours at room temperature.

   Note: AF is a sterile 1X solution containing 0.1 % gelatin (see page XX for ordering information).

2. Using sterile technique in a laminar flow culture hood, completely remove the AF solution from the culture vessel by aspiration.

   Note: It is not necessary to wash the culture surface before adding cells or medium. Coated vessels may be used immediately or wrapped in Parafilm™ sealing film and stored at room temperature for up to 24 hours.

Thaw MEFs

1. Remove the cryovial containing inactivated MEFs from the liquid nitrogen storage tank.
2. Briefly roll the vial between hands to remove frost, and swirl it gently in a 37°C water bath.
3. When only a small ice crystal remains in the vial, remove it from water bath. Spray the outside of the vial with 70% ethanol before placing it in the cell culture hood.
4. Pipet the thawed cells gently into a 15-mL conical tube.
5. Rinse the cryovial with 1 mL of pre-warmed MEF medium. Transfer the medium to the same 15-mL tube containing the cells.
6. Add 4 mL of pre-warmed MEF medium **drop-wise** to the cells. Gently mix by pipetting up and down.

   Note: Adding the medium slowly helps the cells to avoid osmotic shock.

7. Centrifuge the cells at 200 × *g* for 5 minutes.
8. Aspirate the supernatant and resuspend the cell pellet in 5 mL of pre-warmed MEF medium.
9. Remove 20 µL of the cell suspension and determine the viable cell count using your method of choice (e.g., Countess™ Automated Cell Counter).

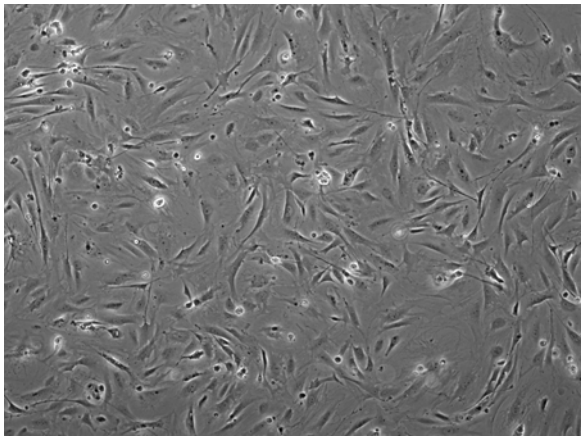
Continued on next page

Plate MEFs

1. Centrifuge the remaining cell suspension (step 9, page 55) at 200 × *g* for 5 minutes at room temperature.
2. Aspirate the supernatant. Resuspend the cell pellet in MEF medium to a density of 2.5×10^5 cells/mL.
3. Aspirate the gelatin solution from the gelatin coated culture vessel.
4. Add the appropriate amount of MEF medium into each culture vessel (refer to the table below).
5. Into each of these culture vessels, add the appropriate amount of MEF suspension (refer to the table below).
   Note: The recommended plating density for Gibco™ Mouse Embryonic Fibroblasts (Irradiated) is 2.5×10^4 cells/cm^2.
6. Move the culture vessels in several quick back-and-forth and side-to-side motions to disperse the cells across the surface of the vessels.
7. Incubate the cells in a 37°C incubator with a humidified atmosphere of 5% CO_2.
8. Use the MEF culture vessels within 3–4 days after plating.

Culture vessel	Surface area	Volume of media	Number of MEFs	Volume of MEF suspension
96-well plate	10 cm^2 per well	0.1 mL	1.0×10^4/well	40 µL
24-well plate	4 cm^2 per well	0.5 mL	5.0×10^4/well	200 µL
12-well plate	2 cm^2 per well	1 mL	1.0×10^5/well	400 µL
6-well plate	10 cm^2	2 mL	2.5×10^5/well	1 mL
60-mm dish	20 cm^2	5 mL	5.0×10^5	2 mL
100-mm dish	60 cm^2	10 mL	1.5×10^6	6 mL
25-cm^2 flask	25 cm^2	5 mL	6.3×10^5	2.5 mL
75-cm^2 flask	75 cm^2	15 mL	1.9×10^6	7.5 mL

Figure 7 Mitotically inactivated Mouse Embryonic Fibroblasts (MEFs) plated on culture vessels coated with Attachment Factor, shown here 4 days after plating.



Coat culture vessels with vitronectin

Vitronectin working concentration

The optimal working concentration of vitronectin is cell line dependent and must be determined empirically. We recommend using a final coating concentration of 0.5 µg/cm^2 on the culture surface. Prior to coating culture vessels, calculate the working concentration of vitronectin using the formula below and dilute the stock appropriately. Refer to the table below for culture surface area and volume required.

$$\text{Working conc.} = \text{Coating conc.} \times \frac{\text{Culture surface area}}{\text{Volume required for surface area}}$$

$$\text{Dilution factor} = \frac{\text{Stock concentration (0.5 mg/mL)}}{\text{Working concentration}}$$

For example, to coat a 6-well plate at a coating concentration of 0.5 µg/cm^2, you will need to prepare 6 mL of diluted vitronectin solution (10 cm^2/well surface area and 1 mL of diluted vitronectin/well; see table below) at the following working concentration:

$$\text{Working concentration.} = 0.5\ \mu g/cm^2 \times \frac{10\ cm^2}{1\ mL} = 5\ \mu g/mL$$

$$\text{Dilution factor} = \frac{0.5\ mg/mL}{5\ \mu g/mL} = 100X \text{ (i.e., 1:100 dilution)}$$

Culture vessel	Surface area	Volume of diluted vitronectin solution
6-well plate	10 cm^2 per well	1.0 mL/well
12-well plate	4 cm^2 per well	0.4 mL/well
24-well plate	2 cm^2 per well	0.2 mL/well
35-mm dish	10 cm^2	1.0 mL
60-mm dish	20 cm^2	2.0 mL
100-mm dish	60 cm^2	6.0 mL
T-25 flask	25 cm^2	2.5 mL
T-75 flask	75 cm^2	7.5 mL

Continued on next page

Coating procedure

Instructions for coating a 6-well culture plate with vitronectin at a coating concentration of 0.5 µg/cm^2 are provided below. For volumes used in other culture vessels, refer to the table on the page 57. To calculate the working concentration of vitronectin used with other coating concentrations and to determine the appropriate dilution factor, use the equations on page 57.

1. Upon receipt, thaw the vial of vitronectin at room temperature and prepare 60-µL aliquots of vitronectin in polypropylene tubes. Freeze the aliquots at –80°C or use immediately.
2. To coat the wells of a 6-well plate, remove a 60-µL aliquot of vitronectin from –80°C storage and thaw at room temperature. You will need one 60-µL aliquot per 6-well plate.
3. Add 60 µL of thawed vitronectin into a 15-mL conical tube containing 6 mL of sterile DPBS without Calcium and Magnesium (Cat. No. 14190) at room temperature. Gently resuspend by pipetting the vitronectin dilution up and down.

   Note: This results in a working concentration of 5 µg/mL (i.e., a 1:100 dilution).
4. Add 1 mL of the diluted vitronectin solution to each well of a 6-well plate (refer to table for the recommended volumes for other culture vessels). When used to coat a 6-well plate (10 cm^2/well) at 1 mL/well, the final concentration will be 0.5 µg/cm^2.
5. Incubate the coated plates at room temperature for 1 hour.

   Note: The culture vessel can now be used or stored at 2–8°C wrapped in laboratory film for up to a week. Do not allow the vessel to dry. Prior to use, pre-warm the culture vessel to room temperature for at least 1 hour.
6. Aspirate the vitronectin solution and discard. It is not necessary to rinse off the culture vessel after the removal of vitronectin. Cells can be passaged directly onto the vitronectin-coated culture vessels.

Coat culture vessels with Geltrex™ matrix

Coating protocol

1. Thaw a 5-mL bottle of Geltrex™ LDEV-Free hESC-Qualified Reduced Growth Factor Basement Membrane Matrix at 2–8°C overnight.
2. For reprogramming experiments using fibroblasts, dilute the thawed Geltrex™ matrix solution 1:1 with cold sterile DMEM/F-12 to prepare 1-mL aliquots (or another volume suitable for your needs) in tubes chilled on ice. These aliquots can be frozen at –20°C or used immediately.

   For reprogramming experiments using hematopoietic progenitor cells (HPCs), such as StemPro™ CD34+ cells, do **not** dilute the Geltrex™ matrix solution 1:1 to create the intermediate dilution. The Geltrex™ matrix solution needs to be twice as strong for reprogramming experiments with HPCs.
3. To create working stocks, dilute an aliquot of Geltrex™ matrix solution 1:50 with cold DMEM on ice. This creates a final dilution of 1:100 for fibroblasts or a final dilution of 1:50 for StemPro™ CD34+ cells.

   Note: An optimal dilution of the Geltrex™ matrix solution may need to be determined for each cell line. Try various dilutions from 1:30 to 1:100.
4. Quickly cover the whole surface of each culture dish with the Geltrex™ matrix solution (see table below).
5. Incubate the dishes in a 37°C, 5% CO_2 incubator for 1 hour.
6. Geltrex™ matrix-coated culture dishes can now be used or stored at 2–8°C for up to a week. Do not allow dishes to dry.
7. Aspirate the diluted Geltrex™ matrix solution from the culture dish and discard. You do not need to rinse off the Geltrex™ matrix solution from the culture dish after removal. Cells can now be passaged directly onto the Geltrex™ matrix-coated culture dish.

Culture vessel	Surface area	Volume of Geltrex™ matrix dilution
6-well plate	10 cm^2/well	1.5 mL/well
12-well plate	4 cm^2/well	750 µL/well
24-well plate	2 cm^2/well	350 µL/well
35-mm dish	10 cm^2	1.5 mL
60-mm dish	20 cm^2	3.0 mL
100-mm dish	60 cm^2	6.0 mL

CytoTune™-EmGFP Reporter control transduction

CytoTune™-EmGFP Sendai Fluorescence Reporter

The CytoTune™-EmGFP Sendai Fluorescence Reporter (Cat. No. A16519), available separately, is a control vector carrying the Emerald Green Fluorescent Protein (EmGFP) gene. The fluorescent control vector allows you to determine whether your cells of interest are amenable or refractive to transduction by Sendai reprogramming vectors. We recommend testing your cell line interest using the CytoTune™-EmGFP Sendai Fluorescence Reporter before starting your reprogramming experiments.

Guidelines for using the CytoTune™-EmGFP Sendai Fluorescence Reporter

- Transducing your cell line of interest using the CytoTune™-EmGFP Sendai Fluorescence Reporter allows you to determine whether or not the cells can be transduced by the Sendai virus vectors; it **does not** indicate the cell line's capability to be reprogrammed.
- Different cell types require different MOIs to express detectable levels of EmGFP. As such, cells should be transduced using a range of different MOIs. We suggest initially transducing your cells with at least 2–3 different MOIs (e.g. 1, 3, and 9).
- Expression of EmGFP should be detectable at 24 hours post-transduction by fluorescence microscopy, and reach maximal levels at 48–72 hours.
- The titer of the CytoTune™ EmGFP vector is lot-dependent. For the specific titer of the vector, refer to the Certificate of Analysis (CoA) available on our website. See **thermofisher.com/cytotune** and search for the CoA by product lot number, which is printed on the vial.
- Avoid re-freezing and thawing of the CytoTune™-EmGFP Sendai Fluorescence Reporter since viral titers can decrease dramatically with each freeze/thaw cycle.

Control transduction protocol for adherent cells

Day –1 to –2: Prepare the cells for transduction

1. 1–2 days before transduction, plate the cells of interest onto the necessary number of wells of a multi-well plate at the appropriate density to achieve 30–60% confluency on the day of transduction (Day 0). One extra well can be used to count cells for viral volume calculations.
2. Culture the cells for one to two more days, ensuring the cells have fully adhered and extended.

Continued on next page

Control transduction protocol for adherent cells, continued

Day 0: Perform transduction

3. On the day of transduction, warm an appropriate volume of cell culture medium for each well to be transduced (e.g., 0.5 mL for each well of a 12-well plate) in a 37°C water bath.
4. Harvest cells from one well of the multi-well plate and perform a cell count. These cells will not be transduced, but will be used to estimate the cell number in the other well(s) plated in Step 1.

   Note: This step is optional and is performed to obtain more accurate MOI calculations. If exact MOIs are not needed, a rough estimate of the number of cells in the well (based on plating density and growth rates) will also suffice.
5. Count (or estimate) the cell number using the desired method (e.g., Countess™ Automated Cell Counter), and calculate the volume of the virus needed to reach the target MOI(s). Titer information can be found on the CoA.

$$\text{Volume of virus (µL)} = \frac{\text{MOI (CIU/cell)} \times \text{number of cells}}{\text{titer of virus (CIU/mL)} \times 10^{-3}\ \text{(mL/µL)}}$$

6. Remove one tube of CytoTune™-EmGFP Sendai Fluorescence Reporter from the –80°C storage. Thaw the vector by first immersing the bottom of the tube in a 37°C water bath for 5–10 seconds, and then removing the tube from the water bath and allowing its contents to thaw at room temperature. Once thawed, briefly centrifuge the tube and place it immediately on ice.
7. Add the calculated volume of CytoTune™-EmGFP Sendai Fluorescence Reporter to the pre-warmed cell culture medium prepared in Step 3. Ensure that the solution is thoroughly mixed by pipetting the mixture gently up and down. Complete the next step within 5 minutes.
8. Aspirate the cell culture medium from the cells, and add the solution prepared in Step 7 to the well. Incubate the cells in a 37°C, 5% CO_2 incubator overnight.

Day 1: Replace medium and culture cells

9. 24 hours after transduction, replace the medium with fresh cell culture medium.

   Note: Depending on your cell type, you should expect to see some cytotoxicity 24–48 hours post-transduction, which can affect >50% of your cells. This is an indication of high uptake of the virus. We recommend that you continue culturing your cells and proceed with the protocol.
10. Visualize the cells on a fluorescence microscope using a standard FITC filter set. EmGFP expression should be visible in some cells (expression will reach maximum levels between 48–72 hours).

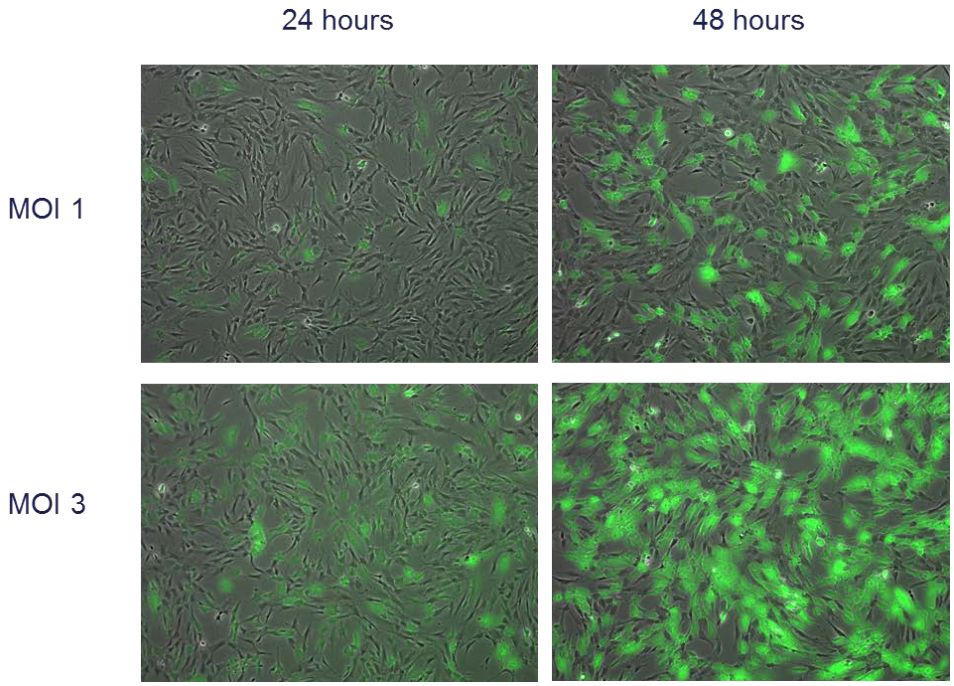
Day 2+: Replace medium and culture cells

11. 48 hours after transduction, replace the medium with fresh cell culture medium.
12. Visualize the cells on a fluorescence microscope using a standard FITC filter set. EmGFP expression should be much brighter than Day 1, and should be visible in many cells (see Figure 8, page 62).

Continued on next page

Expected results

Figure 8 BJ HDFn cells transduced with the CytoTune™-EmGFP Sendai Fluorescence Reporter at the indicated MOI (1 or 3) and at the indicated time post-transduction (24 or 48 hours).



Passage iPSCs with EDTA

Passaging protocol

1. Pre-warm complete Essential 8™ Medium (see page 53) and vitronectin-coated culture vessels (see page 57) to room temperature.
2. Aspirate the spent medium from the vessel containing PSCs and rinse the vessel twice with DPBS without Calcium and Magnesium (refer to the table below for the recommended volume).
3. Add 0.5 mM EDTA in DPBS to the vessel containing PSCs (refer to the table below). Swirl the vessel to coat the entire cell surface.
4. Incubate the vessel at room temperature for 5 to 8 minutes or at 37°C for 4 to 5 minutes. When the cells start to separate and round up, and the colonies appear to have holes in them when viewed under a microscope, they are ready to be removed from the vessel.
5. Aspirate the EDTA solution, and add pre-warmed complete Essential 8™ Medium to the vessel (refer to the table below).
6. Remove the cells from the well(s) by gently squirting medium and pipetting the colonies up. Avoid creating bubbles. Collect cells in a 15-mL conical tube. There may be obvious patches of cells that were not dislodged and left behind. **Do not scrape the cells from the dish in an attempt to recover them.**

   Note: Work with no more than 1 to 3 wells at a time, and work quickly to remove cells after adding Essential 8™ Medium to the well(s), which quickly neutralizes the initial effect of the EDTA. Some cell lines re-adhere very rapidly after medium addition, and must be removed 1 well at a time. Others are slower to re-attach, and may be removed 3 wells at a time.
7. Add an appropriate volume of pre-warmed complete Essential 8™ Medium to each well of a vitronectin-coated 6-well plate so that each well contains 2 mL of medium after the cell suspension has been added. Refer to the table below for the recommended volumes for other culture vessels.
8. Move the vessel in several quick figure eight motions to disperse the cells across the surface of the vessels. Place the vessel gently into the 37°C, 5% CO_2 incubator and incubate the cells overnight.
9. Feed the PSC cells beginning the second day after splitting. Replace the spent medium daily.

   Note: It is normal to see cell debris and small colonies after passage.

Culture vessel	Surface area	DPBS	0.5 mM EDTA in DPBS	Complete Essential 8™ Medium
6-well plate	10 cm^2/well	2 mL/well	1 mL/well	2 mL/well
12-well plate	4 cm^2/well	1 mL/well	0.4 mL/well	1 mL/well
24-well plate	2 cm^2/well	0.5 mL/well	0.2 mL/well	0.5 mL/well
35-mm dish	10 cm^2	2 mL	1 mL	2 mL
60-mm dish	20 cm^2	4 mL	2 mL	4 mL
100-mm dish	60 cm^2	12 mL	6 mL	12 mL

Cryopreserve iPSCs

Freeze iPSCs in iPSC freezing medium

Follow the following protocol to cryopreserve iPSCs maintained on MEF-feeder cells by freezing them in iPSC freezing medium (see page 54 for recipe).

1. Prepare the required volume of fresh iPSC freezing medium and place it on ice (see page 54).
2. Aspirate the culture medium and rinse the dishes twice with D-PBS without Ca^{2+} and Mg^{2+} (2 mL per 35-mm or 4 mL per 60-mm dish).
3. Gently add Collagenase IV solution (page 52) to the culture dish (1 mL per 35-mm or 2 mL per 60-mm dish).
4. Incubate the dish with cells for 5–20 minutes in a 37°C incubator with a humidified atmosphere of 5% CO_2.

   Note: Incubation times may vary among different batches of collagenase. Therefore, the appropriate incubation time should be optimized by examining the colonies periodically under microscope during incubation.
5. Stop the incubation when the edges of the colonies are starting to pull away from the plate.
6. Remove the culture dish from the incubator, aspirate the Collagenase IV solution, and gently rinse the dish with D-PBS without Ca^{2+} and Mg^{2+}.
7. Add 2 mL of iPSC culture medium or DMEM/F-12 and gently dislodge the cells off the surface of the culture dish using a sterile pipette or a cell scraper. Transfer the cells to a sterile 15-mL centrifuge tube. Rinse the dish with additional iPSC medium or DMEM/F-12 to collect any leftover colonies.
8. Centrifuge the cells at 200 × *g* for 2–4 minutes at room temperature.
9. Discard the supernatant, gently tap the tube to dislodge the cell pellet from the tube bottom, and resuspend the cells in **freezing medium A**. After the cell clumps have been uniformly suspended, add an equal volume of **freezing medium B** to the cell suspension in a drop-wise manner while gently swirling the cell suspension to mix.

   Note: At this point, the cells are in contact with DMSO, and work must be performed efficiently with no or minimal delays. After the cells come into contact with DMSO, they should be aliquoted and frozen within 2–3 minutes.
10. Aliquot 1 mL of the cell suspension into each cryovial.
11. Quickly place the cryovials containing the cells in a cryo freezing container (e.g., Mr. Frosty) to freeze the cells at 1°C per minute and transfer them to –80°C overnight.
12. After overnight storage at –80°C, transfer the cells to a liquid nitrogen tank vapor phase for long term storage.

Freeze iPSCs in Essential 8™ freezing medium

Follow the following protocol to cryopreserve iPSCs maintained in Essential 8™ Medium on vitronectin-coated culture dishes by freezing them in Essential 8™ freezing medium (see page 54 for recipe).

1. Prepare the required volume of fresh Essential 8™ freezing medium and place it on ice until use.
   Note: Discard any remaining Essential 8™ freezing medium after use.
2. Aspirate the spent medium from the dish using a Pasteur pipette, and rinse the cells twice with DPBS without Calcium and Magnesium (refer to the table below).
3. Add 0.5 mM EDTA solution to the dish. To adjust the volume of EDTA for various dish sizes, refer to the table below. Swirl the dish to coat the entire cell surface.
4. Incubate the dish at room temperature for 5–8 minutes or at 37°C for 4–5 minutes. When the cells start to separate and round up, and the colonies appear to have holes in them when viewed under a microscope, they are ready to be removed from the vessel.
5. Aspirate the EDTA solution with a Pasteur pipette.
6. Add 1 mL of ice-cold Essential 8™ freezing medium to each well of a 6-well plate. To adjust the volume of Essential 8™ freezing medium for various dish sizes, refer to the table below.
7. Remove the cells by gently squirting the colonies from the well using a 5-mL glass pipette. Avoid creating bubbles. Collect the cells in a 15-mL conical tube and place on ice.
8. Resuspend the cells gently. Aliquot 1 mL of the cell suspension into each cryovial.
9. Quickly place the cryovials containing the cells in a cryo freezing container to freeze the cells at 1°C per minute and transfer them to –80°C overnight.
10. After overnight storage at –80°C, transfer the cells to a liquid nitrogen tank vapor phase for long term storage.

Culture vessel	Surface area	DPBS	0.5 mM EDTA in DPBS	Essential 8™ freezing medium
6-well plate	10 cm^2/well	2 mL/well	1 mL/well	2 mL/well
12-well plate	4 cm^2/well	1 mL/well	0.4 mL/well	1 mL/well
24-well plate	2 cm^2/well	0.5 mL/well	0.2 mL/well	0.5 mL/well
35-mm dish	10 cm^2	2 mL	1 mL	2 mL
60-mm dish	20 cm^2	4 mL	2 mL	4 mL
100-mm dish	60 cm^2	12 mL	6 mL	12 mL

Accessory products

For more information about the following products, refer to our website at or **thermofisher.com** contact Technical Support (page 69).

CytoTune™-iPS products

Product	Quantity	Catalog No.
CytoTune™-iPS 2.0 Sendai Reprogramming Kit	1 × 3 vials	A16517
CytoTune™-iPS 2.0 Sendai Reprogramming Kit (3 Pack)	3 × 3 vials	A16518
CytoTune™-EmGFP Sendai Fluorescence Reporter	1 vial	A16519

Media, sera, and reagents

Product	Quantity	Catalog No.
DMEM with GlutaMAX™-I (high glucose)	500 mL	10569-010
DMEM/F-12, GlutaMAX™ supplement	500 mL	10565018
Essential 8™ Medium	1 kit	A1517001
StemPro™-34 SFM Medium (1X)	500 mL	10639-011
Fetal Bovine Serum (FBS), ES-Cell Qualified	500 mL	16141-079
KnockOut™ Serum Replacement	500 mL	10828-028
Dulbecco's PBS (DPBS) without Calcium and Magnesium	500 mL	14190-144
MEM Non-Essential Amino Acids Solution (10 mM)	100 mL	11140-050
Basic Fibroblast Growth Factor (bFGF)	10 µg	PHG0264
SCF (C-Kit Ligand) Recombinant Human Protein	100 µg	PHC2111
FLT3 Ligand Recombinant Human Protein	10 µg	PHC9414
IL3 Recombinant Human Protein	10 µg	PHC0034
IL6 Recombinant Human Protein	10 µg	PHC0065
GM-CSF Recombinant Human Protein	100 µg	PHC2011
ß-Mercaptoethanol, 55 mM	50 mL	21985-023
GlutaMAX™-I Supplement	100 mL	35050-061
L-Glutamine (200 mM)	100 mL	25030-081
Penicillin-Streptomycin, liquid	100 mL	15140-122

Continued on next page

Accessory products, continued

Cells

Product	Quantity	Catalog No.
Gibco™ Mouse Embryonic Fibroblasts (Irradiated)	1 mL	S1520-100
Human Dermal Fibroblasts, neonatal (HDFn)	1 vial	C-004-5C
StemPro™-34 Medium and CD34+ Cell Kit	1 kit	A14059

Matrices and dissociation reagents

Product	Quantity	Catalog No.
Attachment Factor	100 mL	S-006-100
Vitronectin, truncated human recombinant (VTN-N)	1 mL	A14700
Geltrex™ hESC-qualified Basement Membrane Matrix	5 mL	A14133-02
Collagenase Type IV	1 g	17104-01
0.05% Trypsin/EDTA Solution (1X)	100 mL	25300-054
TrypLE™ Select Cell Dissociation Reagent	100 mL	12563-011
UltraPure™ 0.5 M EDTA, pH 8.0	4 × 100 mL	15575-020
Versene Solution	100 mL	15040-066

Antibodies

Product	Quantity	Catalog No.
TRA-1-60 Mouse anti-human mAb, AlexaFluor™ 488 Conjugate Kit for Live Cell Imaging	1 kit	A25618
TRA-1-60 Mouse anti-human mAb, Alexa Fluor™ 594 Conjugate Kit for Live Cell Imaging	1 kit	A24882
CD44 Rat anti-human/mouse mAb, AlexaFluor™ 488 Conjugate Kit for Live Cell Imaging	1 kit	A25528

Equipment

Product	Quantity	Catalog No.
Countess™ Automated Cell Counter	1 unit	C10227
StemPro™ EZPassage™ Disposable Stem Cell Passaging Tool	10 units	23181-010

Continued on next page

Reagents for RT-PCR

Product	Quantity	Catalog No.
TRIzol™ LS reagent	100 mL	10296-010
SuperScript™ VILO™ cDNA Synthesis Kit	50 reactions	11754-050
AccuPrime™ SuperMix I	200 reactions	12342-010

TaqMan™ Protein Assays

Product	Quantity	Catalog No.
TaqMan™ Protein Assay Kit (hOct3/4)	100 reactions	4405489
TaqMan™ Protein Assay Kit (hSox2)	100 reactions	4405495

Real time RT-PCR

Use the following pre-validated TaqMan™ assays to detect the SeV genome and transgenes in cells reprogrammed using the CytoTune™ 2.0 Sendai reprogramming vectors by real-time RT PCR:

Target	TaqMan™ assay
SeV	Mr04269880_mr
KOS	Mr04421257_mr
Klf4	Mr04421256_mr
cMyc	Mr04269876_mr

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- Product documentation, including:
  – User guides, manuals, and protocols
  – Certificates of Analysis
  – Safety Data Sheets (SDSs; also known as MSDSs)
  Note: For SDSs for reagents and chemicals from other manufacturers, contact the manufacturer.

Limited product warranty

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