1. Immunofluorescence staining

The iPSCs were fixed with 4% paraformaldehyde (PFA), permeabilized with 0.3% Triton X-100, and blocked with 5% bovine serum albumin at room temperature for 1 hour. Cells were incubated with the primary antibody (Table 2) overnight at 4°C, and then visualized with secondary antibodies Alexa 488 and Alexa 555. DNA was stained with DAPI. The stained cells were observed under a confocal microscope (Nikon).

2. Flow cytometry analysis

Cells were collected using Accutase (Gibco), washed twice with phosphate-buffered saline (PBS), and then fixed with 4% PFA. The cells were then permeabilized with 0.3% Triton X-100. Antibodies were diluted as needed (Table 2) and incubated with the cells at room temperature (RT) for 30 minutes. After washing off the primary antibody with PBS, the secondary antibody was incubated with the cells at RT for 15 minutes. Following staining, the cells were washed twice and analyzed using a DxFLEX Flow Cytometer (Beckman Coulter). Unstained cells were used as a negative control to exclude nonspecific fluorescence data.